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Table of Contents

Gover	1
SF 298	2
Introduction	4
Body	4-9
Key Research Accomplishments	9-10
Reportable Outcomes	10
Conclusions	10
References	10-12
Appendices	13-57

INTRODUCTION:

Understanding the mechanism of breast cancer initiation is critical for developing chemoprevention strategies. Normal mammary epithelial cells from which cancer usually originate have limited life span. Immortalization is the first step that leads to continuous growth of mammary epithelial cells (1). Cell cycle regulatory, anti-apoptotic and proapoptotic proteins play a significant role in immortalization process (1). In addition, recent studies suggest that oncogene activation leads to senescence, which serves as an initial barrier for transformation and senescence associated genes are silenced during immortalization (2-5). Immortalized cells attain cancerous growth properties (transformation) due to additional mutations that lead to either loss of tumor suppressor genes or activation of oncogenes. The transcription factor Nuclear factor-kappaB (NFκB) promotes both immortalization and transformation by controlling the expression of cell cycle regulatory, anti-apoptotic and pro-apoptotic genes (6). NF-kB is usually sequestered in the cytoplasm of resting cells through its association with inhibitor-ofkappaB (IkB) proteins and translocates to nucleus upon exposure of cells to cytokines and growth factors (7). NF-kB then binds to response elements and induces the expression of genes involved in invasion, metastasis and chemotherapy resistance. We and others have shown that NF-kB is constitutively active in breast cancer and is responsible for overexpression of several anti-apoptotic genes as well as repression of the pro-apoptotic growth arrest and DNA damage/C/EBP homology (GADD153/CHOP) (8-11). GADD153 is induced when DNA is damaged or cells are under stress. Depending on the extent of damage, cells either repair DNA damage and survive or die. GADD153 is believed to promote death of cells with severe damage, thereby limiting accumulation of cells with mutations (12). Thus, GADD153 is likely to play a role in preventing breast cancer initiation. Because NF-kB reduces GADD153 expression, it is possible that cells that contain constitutively active NF-kB will survive after DNA damage and these cells are more prone for transformation. This award was to test this possibility. Three aims were proposed: 1) To determine whether inhibition of GADD153 by NF-kB is essential for survival and/or transformation of MCF-10A cells when exposed to MMS or grown under nutrient-deprived condition. 2) To determine whether inhibition of GADD153 by NF-kB leads to altered activity of the transcription factor C/EBPβ and differentiation of MCF10A cells. 3) To determine the influence of p53 on the anti-apoptotic function of NF-kB in MCF-10A cells grown under nutrient-deprived condition or exposed to MMS.

BODY:

Specific Aim I:

Task 1:(months 1-5) Establish MCF10A cells overexpressing p65NLS50 or ras and characterize them with respect to constitutive NF-κB activity.

Results: This task is completed. We have established MCF10A cells overexpressing p65NLS50 or Ras (Figure 1A and B). MCF10ApQ and MCF10Aneo cells correspond to parental cells vector alone. MCF10A cells overexpressing p65 (MCF10A/p65 series 1 to 4) showed elevated levels of p65 as well as NF-κB-inducible genes including cyclin D1, Mn-SOD and IκBα (Figure 1A).

<u>Task 2: (Months 6-8) Determine MMS-inducible and nutrient-deprivation inducible expression of GADD153 in MCF10/p65NLS50 cells by Northern and Western blots.</u>
<u>Determine apoptosis by Annexin V labeling, PARP cleavage and DNA laddering.</u>

Results: Cell viability experiments were carried out using the MTT assay and clonogenic assays. MCF-10A/p65NLS50 cells treated with MMS (0 to 10 mM) for 3 to 24 hrs did not show a significant difference in cell viability, as measured by MTT assay compared to vector control cells (data not shown). However, while parental cells (MCF10A/pQ1) were growth inhibited by transforming growth factor beta, MCF-10A/p65(1) cells were resistant (Figure 2A). Unlike in cancer cells, serum starvation did not induce GADD153 in MCF10A cells. Interestingly, MMS-inducible expression of GADD153 was delayed in MCF10A/p65 cells compared to MCF10A/pQ cells (Figure 2B). Thus, NF-κB activation protects mammary epithelial cells against only specific agents.

Task 3: Months (9-15) determine the susceptibility of MCF10A, ras and p65NLS50 cells to MMS-induced transformation. This will be achieved by growing cells in soft agar and matrigel.

Results: MCF10ApQ cells plated on a Matrigel form a highly organized acinar structure. These cells have been used extensively as a model system to study the formation and maintenance of glandular architecture *in vitro*. These acinar structures consist of a hollow lumen surrounded by a single layer of cells, with the hollow lumen formed through a selective apoptosis of centrally located cells (13). Oncogenes such as ErbB2 prevent apoptosis and induce proliferation of the centrally located cells, which leads to the formation of a highly disorganized structure (14). As expected, the parental MCF10A/pQ cells cultured in matrigel formed polarized acinar structures (Figure 3). In contrast, MCF10A/p65 cells formed disorganized structures. Thus, NF-kB activation alone is sufficient for MCF10A cells to show a transformed cell phenotype in matrigel.

Deregulated growth of MCF10Ap65 cells correlates EMT phenotype. When grown in two-dimensional culture, MCF10A/p65 cells exhibited a fibroblastic, spindle-shaped morphology, whereas MCF10A/pQ control cells retained an epithelial-like morphology (Figure 4A). The morphological changes observed in MCF10A/p65 cells resembled that of cells undergoing EMT. To test this possibility, northern and western analyses of epithelial and mesenchymal markers were carried out. MCF10A/p65 cells showed reduced expression levels of epithelial markers, including E-cadherin, Occludin, Desmoplakin and Keratin 18 (K18) compared to MCF10A/pQ control cells. In contrast, expression of the mesenchymal markers Vimentin, and Fibronectin were increased in MCF10A/p65 cells compared to parental cells (Figure 4B).

Increased cellular motility of MCF10A/p65 cells. Increased motility is another hallmark of EMT. Using the *in vitro* wound healing assay, we observed that MCF10A/p65 cells showed a faster rate of motility compared to MCF10A/pQ control cells when monitored over a period of 24 hrs (Figure 5). Taken together, these results confirm NF-κB-dependent EMT of MCF10A cells.

Induction of ZEB-1 and ZEB-2 expression by NF- κ B. To identify downstream targets of NF- κ B involved in induction of EMT phenotype, we used Northern blot and RT-PCR to analyze the expression of transcriptional regulators involved in EMT. MCF10A/p65 cells showed increased expression of the transcriptional repressors ZEB-1 and ZEB-2 (Figure 6A), but not of other transcriptional repressors Snail-1, -2 or -3 (Figure 6B). These results are distinct from that observed in Drosophila, where the NF- κ B homologue dorsal induces EMT through Snail (15). We did not observe a difference in the expression levels of CtBP-1 and CtBP-2 (data not shown), both of which are co-factors required for ZEB-1 and ZEB-2 mediated transcriptional repression (16). IL1- α or TNF α , which induce NF- κ B, increased ZEB-1 and ZEB-2 expression in MCF10A and MDA-MB-231 cells, respectively (Figure 6C).

Overexpression of ZEB-1 induces EMT phenotype in MCF10A cells. To determine whether ZEB-1 alone can induce EMT, MCF10A cells were transduced with control retrovirus (pQCXIN) or with ZEB-1/pQCXIN retrovirus encoding ZEB-1. Two independent clones of MCF10A cells overexpressing ZEB-1 were phenotypically similar to MCF10A/p65 cells (Figure 7A). Northern blot analysis showed higher ZEB-1 and Vimentin, but lower E-cadherin and Desmoplakin levels in MCF10A/ZEB-1 cells compared to control MCF10A/pQN cells. Northern blot analysis was carried out in triplicate for statistical analysis (Figure 7B). These results suggest that ZEB-1 alone can induce the EMT phenotype in MCF10A cells.

Induction of EMT phenotype in MCF10A cells through chronic exposure to TNFα. The experiments described so far demonstrating induction of the EMT phenotype by NF-κB were carried out in an artificial system involving p65 overexpression. To determine whether NF-κB when activated under more physiological conditions could induce the EMT phenotype, MCF10A cells were continuously cultured in medium containing TNFα, a known activator of NF-κB. TNFα did not cause any apparent cytotoxicity to the MCF10A cells and after 22 days of culture, the MCF10A cells showed induction of the EMT phenotype. TNFα-treated cells exhibited a fibroblastic morphology in two-dimensional culture and formed disorganized structures in three-dimensional Matrigel culture, a phenotype reminiscent of that observed with MCF10A/p65 cells (Figure 8).

Reversal of EMT phenotype following removal of TNF α treatment. To determine if the TNF α -induced EMT is reversible, cells treated with TNF α for 22 days were cultured in fresh media without TNF α . Interestingly, the MCF10A cells showed a gradual reversal of the EMT phenotype beginning from 11 days after TNF α removal and acquisition of epithelial morphology by day 18 of TNF α removal (Figure 9A). Reversal of EMT phenotype correlated with corresponding decrease in ZEB-1 and Vimentin expression and increase in E-cadherin and Desmoplakin expression (Figure 9B). Overall, the above experiments involving chronic TNF- α treatment suggest that NF- κ B activation is essential for both initiation and maintenance of EMT phenotype.

Specific aim II:

Task 1: Determine the DNA binding pattern of C/EBPβ and C/EBPβ:GADD153 heterodimers in various cell types by EMSA.

Results: Three isoforms of C/EBPβ are expressed in mammary gland. Normal mammary gland expresses full-length C/EBPβ-1 whereas breast cancers express shorter C/EBPβ-2 (17). C/EBPβ-1 and C/EBPβ-2 are derived from the same transcript utilizing different inframe initiation codons and differ only by 24 amino acids. Overexpression of C/EBPβ-2 alone is sufficient for EMT of MCF-10A cells (18). We did not observe any difference in the mRNA levels of C/EBPβ in MCF10A/pQ and MCF10A/p65 cells (Figure 10A). However, we observed a switch in C/EBPβ isoforms at protein level upon overexpression of p65. While the MCF10A/pQ cells expressed ~50 kDa protein, which corresponds to C/EBPβ-1 isoform, MCF10A/p65 cells expressed C/EBPβ protein of ~46 kDa (Figure 10B). This is most likely C/EBPβ-2 isoform. Overexpression of C/EBPβ-2 isoform has been shown to increase cyclin D (18), which was also observed in MCF10A/p65 cells compared to MCF10A/pQ cells (Figure 10B).

Task 2: Determine the expression levels of C/EBP\$\beta\$ and GADD153-responsive genes in various cell types by transient transfection, Northern and Western blots.

Results: We have measured expression level of β -casein in MCF-10A/pQ and MCF10A/p65 cells. Consistent with changes in C/EBP β protein expression pattern, we observed increased expression of cyclin D1 and osteopontin, C/EBP β responsive genes (Figure 10A and B). Surprisingly, in EMSA, binding of proteins to C/EBP homodimers binding elements as well as to C/EBP/CHOP heterodimer binding elements was reduced in MCF10A/p65 cells compared to parental MCF10A/pQ cells (Figure 10C).

Task 3: Months: Determine the differentiation pathway in various cell types in response to lactogenic stimulation. This will be achieved by measuring β -casein and WAP expression in response to prolactin treatment.

Results: Without any prolactin treatment, we have observed significant change in differentiation program in MCF-10A/p65 cells as outlined below.

In initial studies, MCF10A cells were described as ductal luminal epithelial cells whereas a recent study described MCF10A as myoepithelial cells (19, 20). One explanation for the controversial observation is that MCF10A cells correspond to breast stem cells, which can differentiate into myoepithelial and ductal epithelial cells based on growth condition. Cytokeratin 5 (CK5) is considered as a marker for progenitor/stem cells that can differentiate to either glandular cells (CK8/18+) or myoepithelial cells (SMA+) cells (21). P63, a p53 homologue, is considered as a marker for myoepithelial cells (22). P63 is expressed as several different isoforms; TAp63 and ΔNp63 are the major isoforms derived from alternative promoters. While TAp63 has a transactivation domain and can activate gene expression through p53 response element, ΔNp63 can dominantly inhibit p53 function. ΔNp63 is most commonly expressed isoform in myoepithelial cells (23).

Early passage MCF10A cells (Passage number ~180) obtained from Michigan Cancer Foundation and a late passage cells obtained from our collaborator at our institution were first examined for CK5/6 expression by Western blotting. CK5/6

expression was observed in cells from both sources although expression was substantially lower in early passage cells compared to late passage cells (Figure 11A). In contrast, p63 expression was observed only in late passage cells (Figure 11A). Thus, the late passage cells have acquired some of the characteristics of myoepithelial cells. Most of the studies described below were done using late passage cells because myoepithelial cells have been shown to be replaced by cells that have converted from epithelial to mesenchymal phenotype when breast cancer progresses to invasive phenotype (24).

Because MCF10A/p65 cells showed fibroblastic phenotype compared to MCF10A/pQ cells, we examined the effect of p65 on p63 expression by Western blotting. The p63 expression was observed in MCF-10A/pQ cells but not MCF-10A/p65 cells (Figure 11C), which suggests that NF-κB converts myoepithelial cells to myofibroblasts. Northern analysis for p63 further confirmed the results of Western analysis (Figure 11B and C)). MCF-10A cells exposed to TNF, IL-1 and TPA showed reduced levels of ΔNp63, which suggests NF-κB-dependent repression of ΔNp63 (Figure 11D). ZEB-1 may mediate NF-κB-mediated repression of p63 because p63 levels were lower in MCF-10A cells overexpressing ZEB-1 compared to parental cells (Figure 11E). These results clearly suggest that NF-κB can alter differentiation of myoepithelial cells without the requirement of any costimulus.

Specific aim III

Task 1: Establish cells overexpressing p53 dominant-negative mutants and characterize them for p53 activity. Stable cell lines overexpressing p53V144A will be established.

Results: We generated MCF-10A cells overexpressing dominant negative p53 (Figure 12B). Unfortunately, this particular variant of p53 failed to have a dominant negative affect on wild type p53 of MCF10A cells because doxorubicin-inducible activation of p53-responsive gene p21 was same in parental and dominant negative p53 overexpressing cells (Figure 12A). Therefore, we used retrovirus vectors with shRNA against p53 from Open Biosystems to generate MCF-10A cells lacking p53. However, this approach also proved not useful in generating MCF10A cells with reduced p53 (Figure 12B). We next examined whether p53 expression can be reduced by transient expression of p53 siRNA. Although p53 siRNA reduced p53 protein levels (Figure 12B), transfection reagent itself proved toxic to cells and appears to activate p53 because we observed higher levels of p53 in cells transfected with irrelevant luciferase siRNA.

Task 2: Determine MMS sensitivity, NF-κB activity and GADD153 expression in various cell types.

Result: This task could not be completed because of our inability to develop MCF10A cells lacking p53 activity

<u>Task 3: Determine the MMS-induced transformation frequency of cells that express p53 dominant negative mutant.</u>

Results: This task could not be completed because of our inability to develop MCF10A cells lacking p53.

Mechanism of GADD153 repression by NF-κB: We have performed several mechanistic studies to understand repression of GADD153 by p65:

- a) Exon1 of GADD153 contains NF-κB repressible element: Analysis of a series of deletion mutants of GADD153 promoter-CAT reporter revealed the presence of NF-κB-repressible element in exon 1 of GADD153 gene (Figure 13A). The activity of the mutant lacking exon 1 (pG3mut1/CAT) was not repressed by p65 compared to wild type reporter (pG3a/CAT). Cloning of Exon 1 upstream of the GADD153 promoter (pa91pG3mut1/CAT) or downstream of the CAT reporter (pb pG3mut1/CAT) restored p65-mediated inhibition of CAT activity (Figure 13B). Exon 1 contains a transferable p65 repressible element as an RSV promoter/CAT reporter with exon 1 of GADD153 but not unrelated 91 base long sequence was repressed by p65 (Figure 13A). Further deletion analysis of Exon 1 revealed that +41 to +71 bp region of Exon 1 contains p65-repressible element (Figure 13C).
- b) Transcriptional activity of p65 is required for repression of GADD153: A mutant of p65 with has reduced transactivation potential due to mutation of protein kinase A phosphorylation site (S276A) was less efficient in repressing GADD153 expression compared to wild type p65 (Figure 14). These results suggest that a protein whose expression is induced by p65 is responsible for the repression of GADD153 expression.
- c) Gfi family members are involved in repression of GADD153: The 30 bp region in exon 1 of GADD153 contains a putative binding site for the Gfi repressor family. The members in this family include Snail, Slug, Gfi-1, Gfi-1B, IA-1 and Mlt1(25, 26). Most of these proteins function as transcription repressors. Electrophoretic mobility shift assay (EMSA) showed specific binding of Snail-1 and Gfi-1 to the 91 bp region exon 1 of GADD153 (Figure 15A and B). We also observed binding of Gfi-1 to GADD153 promoter in MDA-MB-231 cells by chromatin immunoprecipitation assay (Figure 16). In transient transfection assays, Gfi-1 repressed GADD153 expression (Figure 17). Snail is a NF-κB inducible gene (27), although we did not observe NF-κB-dependent induction of Snail in both MCF10A and MDA-MB-231 cells (data not shown). Transcription factor binding motif search revealed the presence of 1, 6 and 2 putative NF-κB binding sites in Gfi-1, Mlt1 and IA-1 genes, respectively. Although our initial studies suggested that Gfi1 is NF-κB-inducible, we could not reproduce this results in several other cell types. We are currently investigating whether Mlt1 and IA-1 are induced by NF-κB, which play a role in repression of GADD153.

KEY RESEARCH ACCOMPLISHMENTS:

- MCF10A subtypes with myoepithelial characteristics overexpressing p65 subunit of NF-κB show an EMT phenotype. EMT phenotype correlated with loss of expression of myoepithelial cell marker ΔNp63.
- EMT correlates with NF-kB-dependent expression of EMT-associated transcription repressors ZEB-1 and ZEB-2. ZEB-1 alone could induce EMT.
- Chronic exposure of MCF10A cells to TNFα, an inducer of NF-κB, also induced EMT phenotype.

- MCF10A cells overexpressing p65 show altered expression pattern of C/EBPβ isoforms.
- The exon 1 of GADD153 contains NF-κB repressible element. NF-κB repressible element binds to Snail/Gfi family transcription repressors. Snail/Gfi family transcription repressors may play a role in NF-κB-mediated repression of GADD153.

Reportable outcomes: Three abstracts have been published and two manuscripts will be submitted in one month. Part of the data is also reported in a review written by the PI.

Review article: NF-κB and breast cancer. Current Problems in Cancer 26:277-310 (2002)

Abstracts:

- 1) Hui Lin Chua, Sunil Badve and **Harikrishna Nakshatri**. NF-κB induces epithelial to mesenchymal transition phenotype to immortalized mammary epithelial cells. NF-κB: from bench to bedside. Keystone symposia, Snow Bird, Utah (January 10-15, 2004).
- 2) An NF-kB repressible element in the 5' untranslated region of the pro-apoptotic GADD153/CHOP gene. Hui Lin Chua and Harikrishna Nakshatri. American Association for Cancer Research 94th Annual Meeting Washington, DC, June 10-14, 2003.
- 3) Hui Lin Chua, Sunil Badve and Harikrishna Nakshatri. NF-kappaB promotes epithelial-to-mesenchymal transition of immortalized mammary epithelial cells through ZEB-1 and ZEB-2. Era of Hope, Department of Defense Breast Cancer Research Program Meeting, Philadelphia, June 8-11, 2005.

Conclusions: NF- κ B can influence breast cancer progression by multiple mechanisms. This study showed a role for NF- κ B in converting myoepithelial cells to myofibroblasts. Myofibroblasts constitute important stromal cells that provide secretory factors required for invasion and metastasis of breast cancer (28). We have also identified a nexus between NF- κ B and Snail/Gfi family of transcription factors. While transcription activation by NF- κ B is very well studied, transcription repression by NF- κ B is not known. Because Snail/Gfi family members are transcription repressors, they may mediate transcription repression by NF- κ B.

References:

- 1. Wright WE, Shay JW. Telomere dynamics in cancer progression and prevention: fundamental differences in human and mouse telomere biology. Nat Med 2000;6(8):849-51.
- 2. Michaloglou C, Vredeveld LC, Soengas MS, et al. BRAFE600-associated senescence-like cell cycle arrest of human naevi. Nature 2005;436(7051):720-4.
- 3. Braig M, Lee S, Loddenkemper C, et al. Oncogene-induced senescence as an initial barrier in lymphoma development. Nature 2005;436(7051):660-5.

- 4. Chen Z, Trotman LC, Shaffer D, et al. Crucial role of p53-dependent cellular senescence in suppression of Pten-deficient tumorigenesis. Nature 2005;436(7051):725-30.
- 5. Collado M, Gil J, Efeyan A, et al. Tumour biology: senescence in premalignant tumours. Nature 2005;436(7051):642.
- 6. Ghosh S, Karin M. Missing pieces in the NF-kappaB puzzle. Cell 2002;109 Suppl:S81-96.
- 7. Karin M, Cao Y, Greten FR, Li ZW. NF-kappaB in cancer: from innocent bystander to major culprit. Nature Rev Cancer 2002;2(4):301-10.
- 8. Nakshatri H, Bhat-Nakshatri P, Martin DA, Goulet RJ, Jr., Sledge GW, Jr. Constitutive activation of NF-kappaB during progression of breast cancer to hormone-independent growth. Mol Cell Biol 1997;17(7):3629-39.
- 9. Sovak MA, Bellas RE, Kim DW, et al. Aberrant nuclear factor-kappaB/Rel expression and the pathogenesis of breast cancer. J Clin Invest 1997;100(12):2952-60.
- 10. Cogswell PC, Guttridge DC, Funkhouser WK, Baldwin Jr AS. Selective activation of NF-kappaB subunits in human breast cancer: potential roles for NF-kappaB2/p52 and for Bcl-3. Oncogene 2000;19(9):1123-31.
- 11. Nozaki S, Sledge Jr GW, Nakshatri H. Repression of GADD153/CHOP by NF-kappaB: a possible cellular defense against endoplasmic reticulum stress-induced cell death. Oncogene 2001;20(17):2178-85.
- 12. Kaufman RJ. Stress signaling from the lumen of the endoplasmic reticulum: coordination of gene transcriptional and translational controls. Genes Dev 1999;13(10):1211-33.
- 13. Mills KR, Reginato M, Debnath J, Queenan B, Brugge JS. Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) is required for induction of autophagy during lumen formation in vitro. Proc Natl Acad Sci U S A 2004;101(10):3438-43.
- 14. Debnath J, Mills KR, Collins NL, Reginato MJ, Muthuswamy SK, Brugge JS. The role of apoptosis in creating and maintaining luminal space within normal and oncogene-expressing mammary acini. Cell 2002;111(1):29-40.
- 15. Akimaru H, Hou DX, Ishii S. Drosophila CBP is required for dorsal-dependent twist gene expression. Nat Genet 1997;17(2):211-4.
- 16. Postigo AA, Dean DC. Differential expression and function of members of the zfh-1 family of zinc finger/homeodomain repressors. Proc Natl Acad Sci U S A 2000;97(12):6391-6.
- 17. Eaton EM, Hanlon M, Bundy L, Sealy L. Characterization of C/EBPbeta isoforms in normal versus neoplastic mammary epithelial cells. J Cell Physiol 2001;189(1):91-105.
- 18. Bundy LM, Sealy L. CCAAT/enhancer binding protein beta (C/EBPbeta)-2 transforms normal mammary epithelial cells and induces epithelial to mesenchymal transition in culture. Oncogene 2003;22(6):869-83.
- 19. Gordon LA, Mulligan KT, Maxwell-Jones H, Adams M, Walker RA, Jones JL. Breast cell invasive potential relates to the myoepithelial phenotype. Int J Cancer 2003;106(1):8-16.
- 20. Tait L, Soule HD, Russo J. Ultrastructural and immunocytochemical characterization of an immortalized human breast epithelial cell line, MCF-10. Cancer Res 1990;50(18):6087-94.

- 21. Boecker W, Buerger H. Evidence of progenitor cells of glandular and myoepithelial cell lineages in the human adult female breast epithelium: a new progenitor (adult stem) cell concept. Cell Prolif 2003;36 Suppl 1:73-84.
- 22. Westfall MD, Pietenpol JA. p63: molecular complexity in development and cancer. Carcinogenesis 2004.
- 23. Barbareschi M, Pecciarini L, Cangi MG, et al. p63, a p53 homologue, is a selective nuclear marker of myoepithelial cells of the human breast. Am J Surg Pathol 2001;25(8):1054-60.
- 24. Rizki A, Bissell MJ. Homeostasis in the breast: it takes a village. Cancer Cell 2004;6(1):1-2.
- 25. Tateno M, Fukunishi Y, Komatsu S, et al. Identification of a novel member of the snail/Gfi-1 repressor family, mlt 1, which is methylated and silenced in liver tumors of SV40 T antigen transgenic mice. Cancer Res 2001;61(3):1144-53.
- 26. Goto Y, De Silva MG, Toscani A, Prabhakar BS, Notkins AL, Lan MS. A novel human insulinoma-associated cDNA, IA-1, encodes a protein with "zinc-finger" DNA-binding motifs. J Biol Chem 1992;267(21):15252-7.
- 27. Bachelder RE, Yoon SO, Franci C, de Herreros AG, Mercurio AM. Glycogen synthase kinase-3 is an endogenous inhibitor of Snail transcription: implications for the epithelial-mesenchymal transition. J Cell Biol 2005;168(1):29-33.
- 28. Allinen M, Beroukhim R, Cai L, et al. Molecular characterization of the tumor microenvironment in breast cancer. Cancer Cell 2004;6(1):17-32.

- Figure 1: A) Generation of MCF10A cells overexpressing p65NLS50. MCF10A cells were infected with control retrovirus (pQ series) or p65NLS coding retrovirus (p65 1-4). Four independent mass cultures after selection with G418 were analyzed for expression of p65 and NF-κB inducible genes by Western blotting. B) Ras expression in parental (MCF10A/neo) cells and ras overexpressing cells (MCF10A/ras). Lysate from a mammary tumor developed in a MMTV-Ras transgenic mice (MMT-11ras) was used as a positive control.
- Figure 2: A) Sensitivity of parental (MCF10A/pQ1) and p65 overexpressing cells (MCF10A/p65) to TGFβ1. Cells were treated with TGFβ1 for six days and cell growth was measured by MTT assay. Note that MCF10A/p65 cells are resistant to TGFβ1. B) MMS inducible expression of GADD153 in MCF10A/pQ and MCF10A/p65 cells. Cells were treated with 500 micromolar MMS for indicated time and GADD153 expression was measured by Northern blotting. Note differences in GADD153 induction between two cell types.
- Figure 3: MCF10A/p55 cells form a highly disorganized structure compared to MCF10A/pQ cells, which form acini-like structure, upon culturing in matrigel.
- Figure 4A: MCF10A/p65 cells show fibroblast morphology compared to MCF10A/pQ cells, which show epithelial morphology when grown on plastic.
- Figure 4B: MCF10A/p65 cells show lower expression of epithelial markers (E-cadherin, K18, Desmoplakin) and increased mesenchymal markers (Fibronectin and Vimentin) compared to MCF10A/pQ cells. E-cadherin expression was measured by both Northern (left) and Western blots (right) whereas expression of other genes was measured by Northern blotting.
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- Figure 6: MCF10A/p65 cells overexpress ZEB-1 and ZEB-2 compared to MCF10A/pQ cells. ZEB-1 and ZEB-2 expression were measured by Northern blotting. B) p65 overexpression had no affect on Snail-1, Snail-2 and Snail-3 expression. Snail expression was measured by RT-PCR. C) TNFα and IL-1α, potent NF-κB activators, induce ZEB-1 and ZEB-2 in MCF-10A and MD231 cells, respectively. Expression of ZEB-1 and ZEB-2 was measured by Northern blotting.

- Figure 7A: MCF10A cells overexpressing ZEB-1 show fibroblastic morphology.
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- Figure 9B: Epithelial (E-cadherin and Desmoplakin), mesenchymal (Vimentin) and ZEB-1 expression in untreated cells (no Tx), TNF α treated cells (TNF α +) and cells treated with TNF α for 22 days but withdrawn subsequently for indicated days (TNF α -) was measured by Northern blotting. Note that in TNF α cells ZEB-1 and Vimentin expression is reduced but E-cadherin and Desmoplakin expression is increased compared to TNF α + cells.
- Figure 10: A) P65 overexpression does not affect the level of C/EBPβ mRNA but increases osteopontin expression. C/EBPβ and osteopontin expression were measured by Northern analysis. MCF10A/pQ(A) cells correspond to control cells whereas MCF10A/p65 (A, B, J) correspond to cells overexpressing p65. B) MCF10A/p65 cells contain a unique form of C/EBPβ protein compared to MCF10A/pQ cells as measured by Western blotting.
- Figure 10C: Transcription factor binding to an oligonucleotide with consensus binding sequence for C/EBP homodimer (C/EBP-C/EBP) or C/EBP/GADD153 heterodimer (C/EBP-GADD153) was determined by electrophoretic mobility shift assays using nuclear extracts from MCF10A/pQ and MCF10A/p65 cells. Oligonucleotide with a binding site for SP-1 was used as a control. Note the effect of p65 overexpression on transcription factor binding to C/EBP-C/EBP and C/EBP-GADD153 elements.
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- Figure 12: A) Generation of MCF10A cells overexpressing dominant negative mutant of p53 (p53dnV143A). However, dominant negative mutant had no effect on doxorubicin-inducible expression of p53 target gene p21. Expression of p53 and p21 was determined by Western blotting. B) p53 levels are reduced in MCF10A treated transiently with siRNA against p53 but not in cells stably expressing shRNA (00546) against p53. P53 expression was measured by Western blotting.
- Figure 13A: Exon 1 of GADD153 promoter contains p65 repressible element. Cos-1 cells were transfected with indicated CAT reporters (5 micrograms), p65 or p50 expression vectors (0.5 micrograms) and 2 micrograms of β-galactosidase expression vector pCH110. CAT activity was measured 48 hour after transfection. The pG3a/CAT correspond to wild type GADD153 promoter with sequences -247 to +91 (which includes exon 1). The pG3mut1/CAT contains -247 to +1 sequence of GADD153 promoter. Wild type RSV/CAT and RSV/CAT with exon 1 of GADD 153 (p91/CAT) or irrelevant 91 bases (pcontr91/CAT) is also shown. Note that p65 represses pG3a/CAT but not pG3mut1/CAT.
- Figure 13B: Insertion of exon 1 upstream of -247 to +1 of GADD153 sequence (pa91 G3mut/CAT) or downstream of CAT gene (pb91 pG3mut1/CAT) in pG3mut1/CAT restores inhibitory effect of p65.
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- Figure 14: The p65 with mutation of S276 to alanine [p65(S276mut)] was less efficient than wild type p65 in repressing GADD153 promoter. P5W1/CAT corresponds to CAT reporter with -954 to +91 bases of GADD153 promoter. Note that p65S276mut is less efficient than wild type p65 in activating NF κ B/CAT reporter.
- Figure 15: A) The exon 1 of GADD153 binds to Snail-1. Cos-1 cells were transfected with control plasmid pCMV2 or Flag-epitope tagged Snail-1. Radiolabeled 91 bp-untranslated region of exon 1 of GADD153 was incubated with nuclear extract from transfected cells and DNA: protein complex was resolved on an acrylamide gel. A specific DNA: protein complex was observed with nuclear extract from Snail-1 transfected cells. Flag antibody disrupted this complex. B) Binding of Gfi-1 to exon 1 of GADD153. Experiments were done as in A. Gfi-1:DNA complex was detectable with nuclear extracts from cells transfected with Gfi-1. This complex was disrupted by Gfi-1 specific but not isotype antibody. Also Gfi-1 binding was reduced substantially when exon-1 with Gfi-1 binding site mutation was used as a probe (p91mutb).

- Figure 16: Binding of endogenous Gfi-1 to GADD153 promoter was determined by chromatin immunoprecipitation assay. MD435 cells were used because these cells express higher levels of Gfi-1. Gfi-1 immunoprecipitates from cross-linked cells contain sequences corresponding to exon 1 but exon 4 of GADD153. Gfi-1 binding was higher in cells that are overexpressing Gfi-1 (MD435/Gfi-1) compared to cells transfected with vector (MD435/pQN). PCR amplification of input DNA is also shown.
- Figure 17: Gfi-1 represses GADD153 promoter. Cos-1 cells were transfected with pG3/CAT or pG3/CAT with mutation of Gfi-1 binding site (pG3mutb/CAT) and p65 or Gfi-1 expression vector. PCH110 was used as an internal control to measure transfection efficiency. pG3a/CAT but not pG3mutb/CAT activity was lower in cells transfected with p65 or Gfi-1. Thus, Gfi-1 functions as repressor of GADD153.

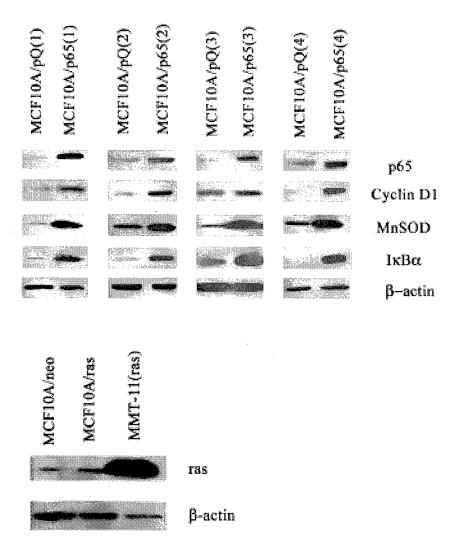
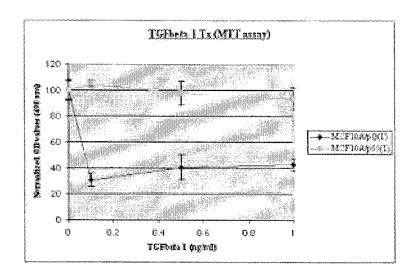


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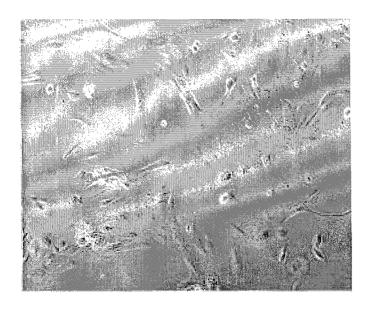
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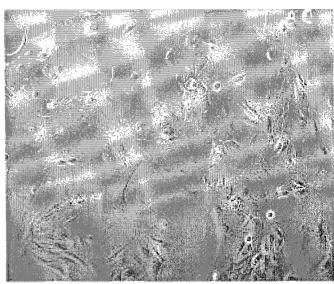
MCF10A/pQ(1) MCF10A/pQ(4) MCF10A/p65(1) MCF10A/p65(4)

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MCF10A/pQ(1)

MCF10A/pQ(4)





MCF10A/p65(1)

MCF10A/p65(4)



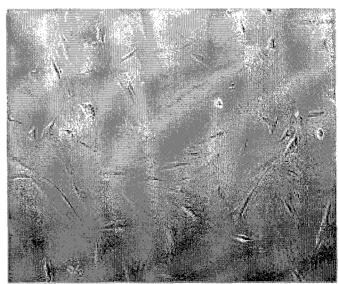


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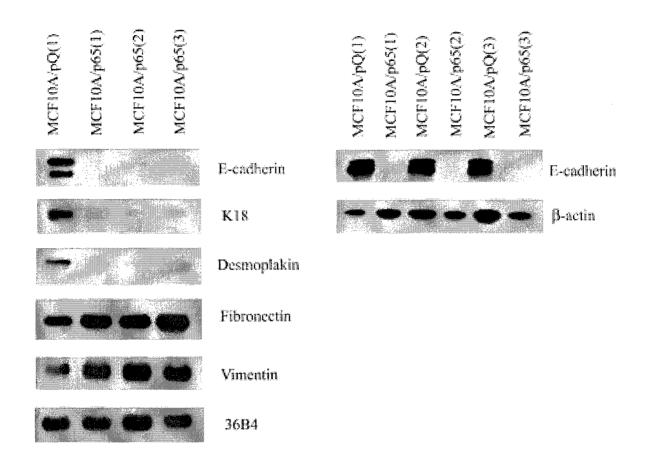


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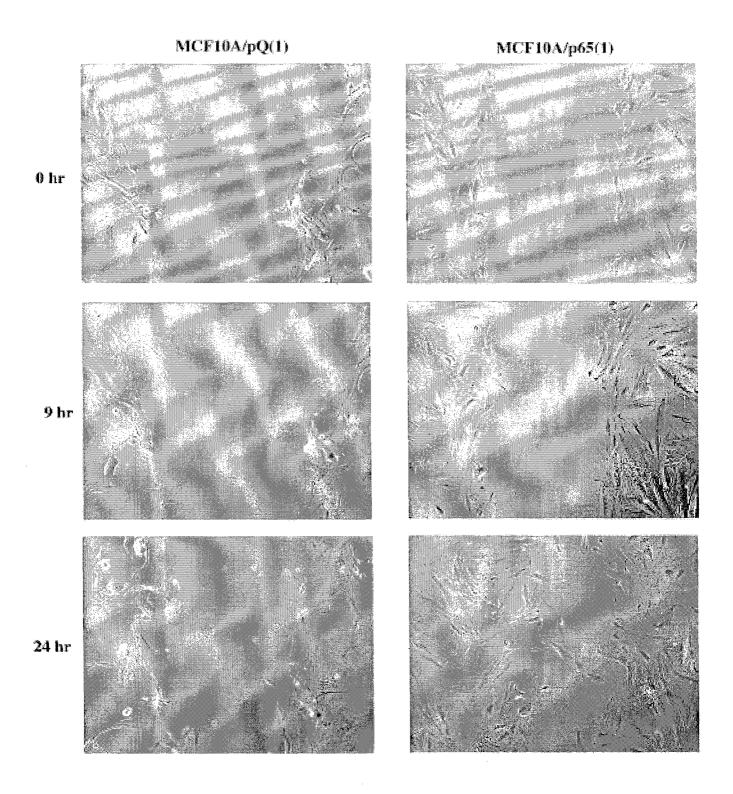
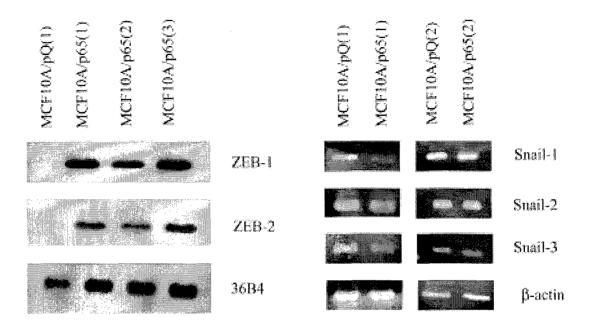


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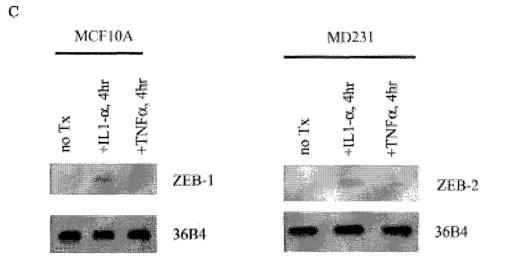
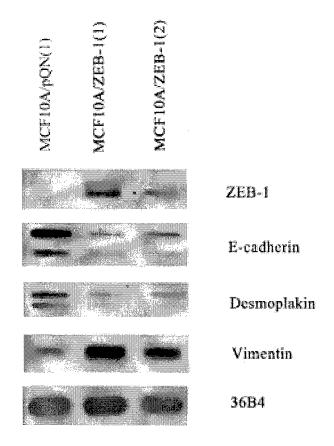


Figure 6: MCF10A/p65 cells overexpress ZEB-1 and ZEB-2 compared to MCF10A/pQ cells. ZEB-1 and ZEB-2 expression were measured by Northern blotting. B) p65 overexpression had no affect on Snail-1, Snail-2 and Snail-3 expression. Snail expression was measured by RT-PCR. C) TNFα and IL-1α, potent NF-κB activators, induce ZEB-1 and ZEB-2 in MCF-10A and MD231 cells, respectively. Expression of ZEB-1 and ZEB-2 was measured by Northern blotting.

MCF10A/pQN(1) MCF10A/pQN(2) MCF10A/ZEB1(1) MCF10A/ZEB1(2)

Figure 7A: MCF10A cells overexpressing ZEB-1 show fibroblastic morphology.



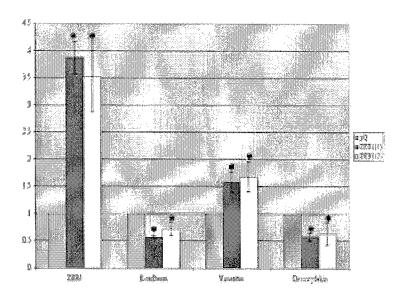


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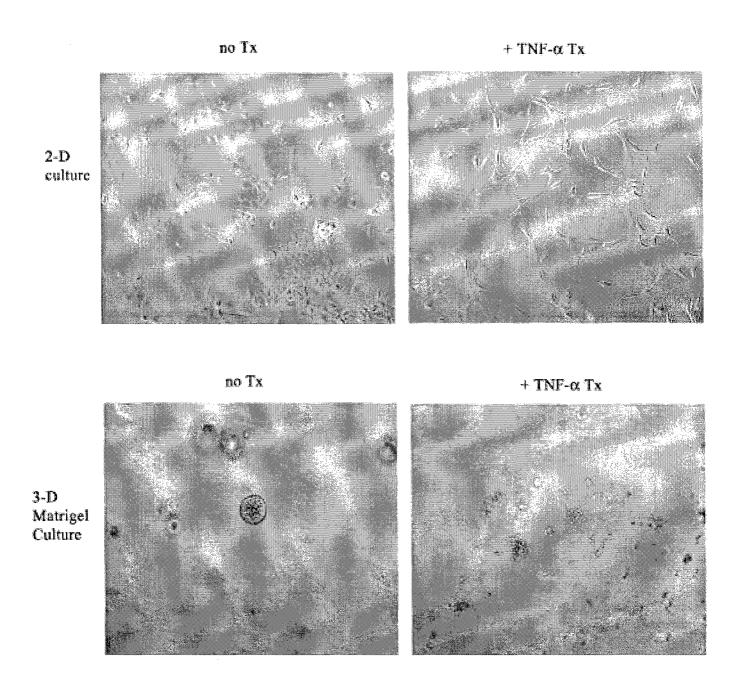


Figure 8: MCF10A cells exposed to TNFa for 22 days show fibroblastic morphology on plastic (top panel) and form a disorganized structure in matrigel (bottom panel).

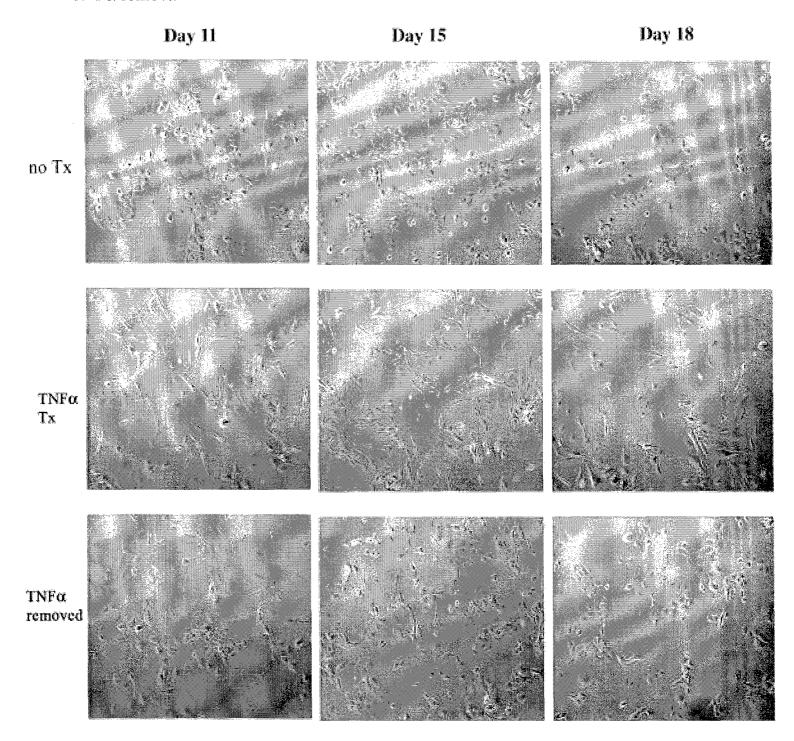


Figure 9A: The effect of TNFα on MCF10A is reversible. Cells were treated with TNFα for 22 days and grown subsequently without TNFα (bottom panel) or with TNFα (middle panel) for indicated days. Untreated cells are shown in the top panel.

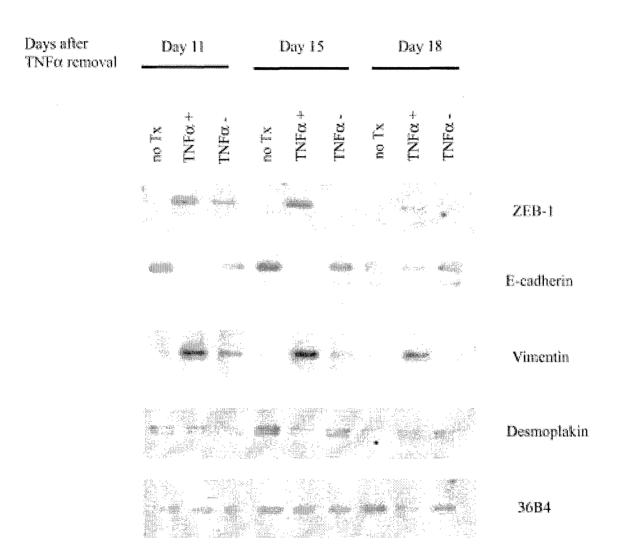


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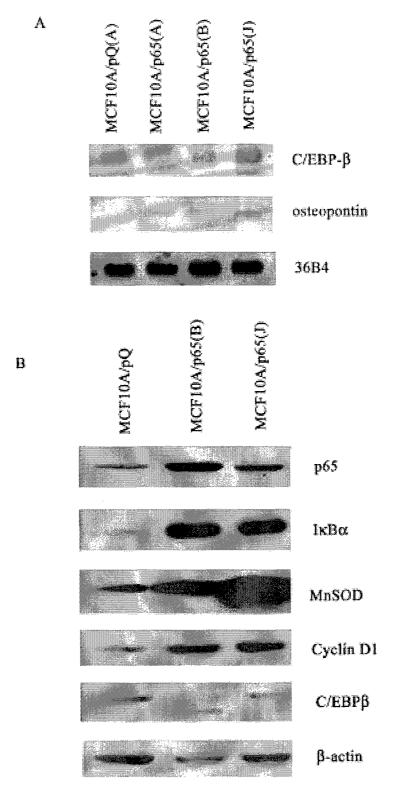


Figure 10: A) P65 overexpression does not affect the level of C/EBPβ mRNA but increases osteopontin expression. C/EBPβ and osteopontin expression were measured by Northern analysis. MCF10A/pQ(A) cells correspond to control cells whereas MCF10A/p65 (A, B, J) correspond to cells overexpressing p65. B) MCF10A/p65 cells contain a unique form of C/EBPβ protein compared to MCF10A/pQ cells as measured by Western blotting.

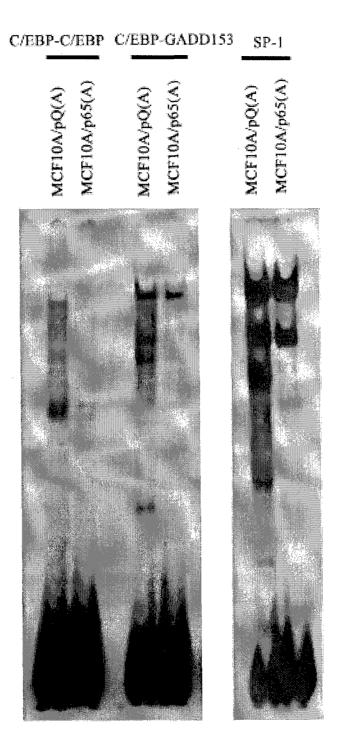


Figure 10C: Transcription factor binding to an oligonucleotide with consensus binding sequence for C/EBP homodimer (C/EBP-C/EBP) or C/EBP/GADD153 heterodimer (C/EBP-GADD153) was determined by electrophoretic mobility shift assays using nuclear extracts from MCF10A/pQ and MCF10A/p65 cells. Oligonucleotide with a binding site for SP-1 was used as a control. Note the effect of p65 overexpression on transcription factor binding to C/EBP-C/EBP and C/EBP-GADD153 elements.

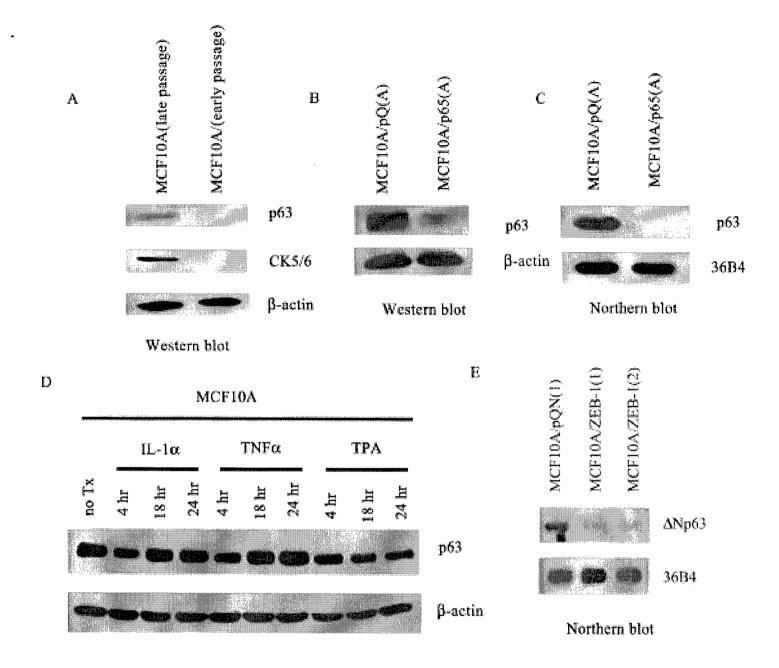


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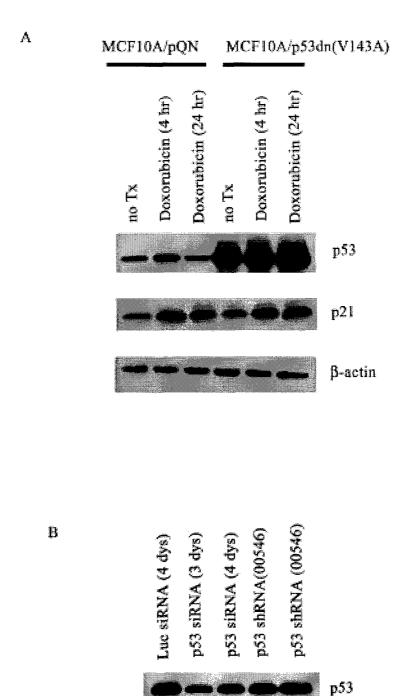
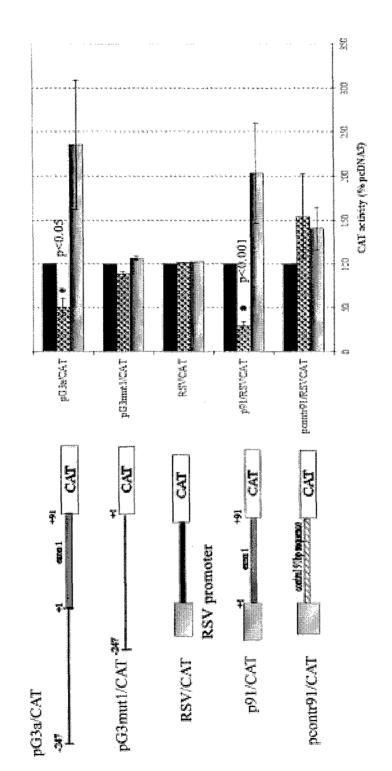


Figure 12: A) Generation of MCF10A cells overexpressing dominant negative mutant of p53 (p53dnV143A). However, dominant negative mutant had no effect on doxorubicin-inducible expression of p53 target gene p21. Expression of p53 and p21 was determined by Western blotting. B) p53 levels are reduced in MCF10A treated transiently with siRNA against p53 but not in cells stably expressing shRNA (00546) against p53. P53 expression was measured by Western blotting.





GADD 153 (p91/CAT) or irrelevant 91 bases (pcontr91/CAT) is also shown. Note Figure 13A: Exon 1 of GADD153 promoter contains p65 repressible element. Cos-1 cells expression vector pCH110, CAT activity was measured 48 hour after transfection. The pG3a/CAT correspond to wild type GADD153 promoter with sequences -247 to +91 (which includes exon 1). The pG3mut1/CAT contains -247 to +1 sequence of GADD153 promoter. Wild type RSV/CAT and RSV/CAT with exon 1 of were transfected with indicated CAT reporters (5 micrograms), p65 or p50 expression vectors (0.5 micrograms) and 2 micrograms of β-galactosidase that p65 represses pG3a/CAT but not pG3mut1/CAT.

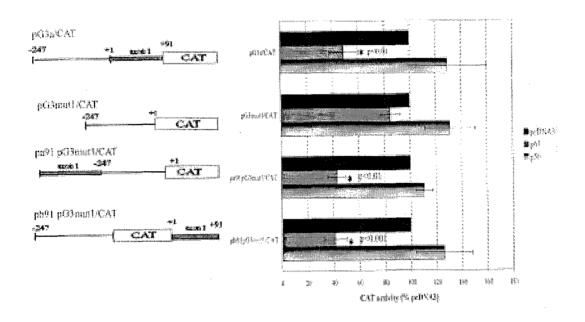


Figure 13B: Insertion of exon 1 upstream of -247 to +1 of GADD153 sequence (pa91 G3mut/CAT) or downstream of CAT gene (pb91 pG3mut1/CAT) in pG3mut1/CAT restores inhibitory effect of p65.

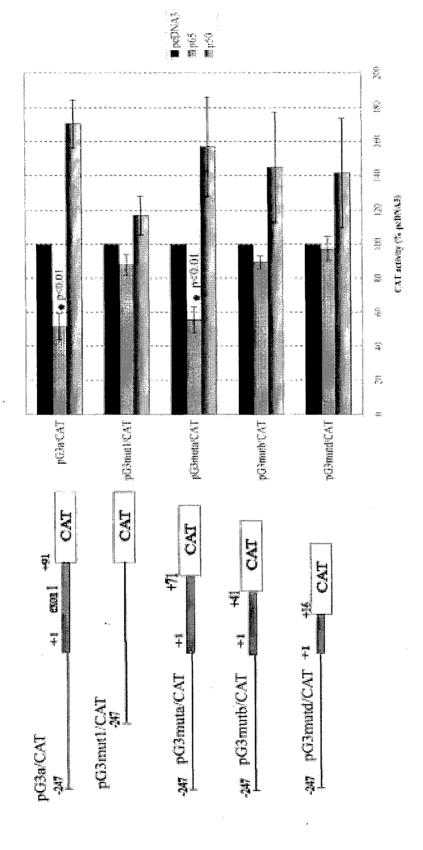


Figure 13C; pG3a/CAT reporter lacking sequences from +42 to +71 of GADD 153 (pG3muth/CAT) was not repressed by p65 whereas pG3a/CAT reporter with +1 to +71 (pG3muta/CAT) was repressed by p65.

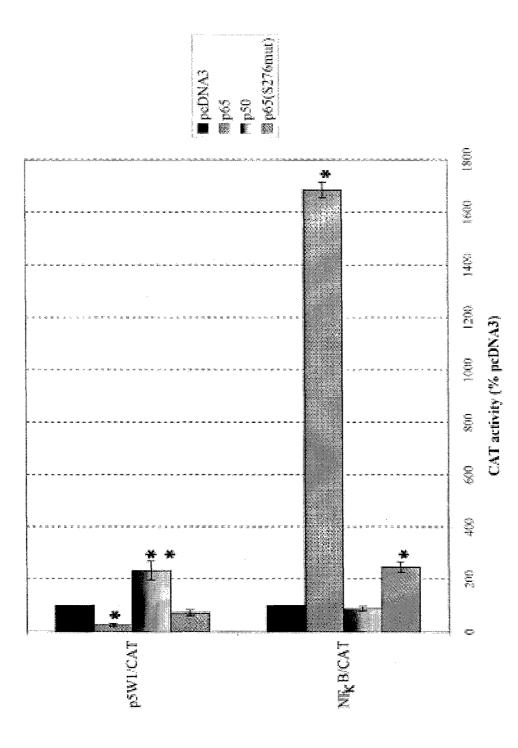


Figure 14: The p65 with mutation of S276 to alanine [p65(S276mut)] was less efficient than wild type p65 in repressing GADD153 promoter. P5W1/CAT corresponds to CAT reporter with -954 to +91 bases of GADD153 promoter. Note that p65S276mut is less efficient than wild type p65 in activating NFkB/CAT reporter.

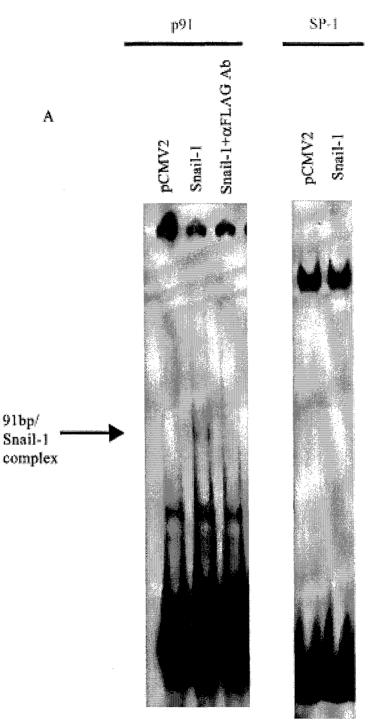


Figure 15 A The exon 1 of GADD153 binds to Snail-1. Cos-1 cells were transfected with control plasmid pCMV2 or Flag-epitope tagged Snail-1. Radiolabeled 91 bp-untranslated region of exon 1 of GADD153 was incubated with nuclear extract from transfected cells and DNA; protein complex was resolved on an acrylamide gel. A specific DNA; protein complex was observed with nuclear extract from Snail-1 transfected cells. Flag antibody disrupted this complex.

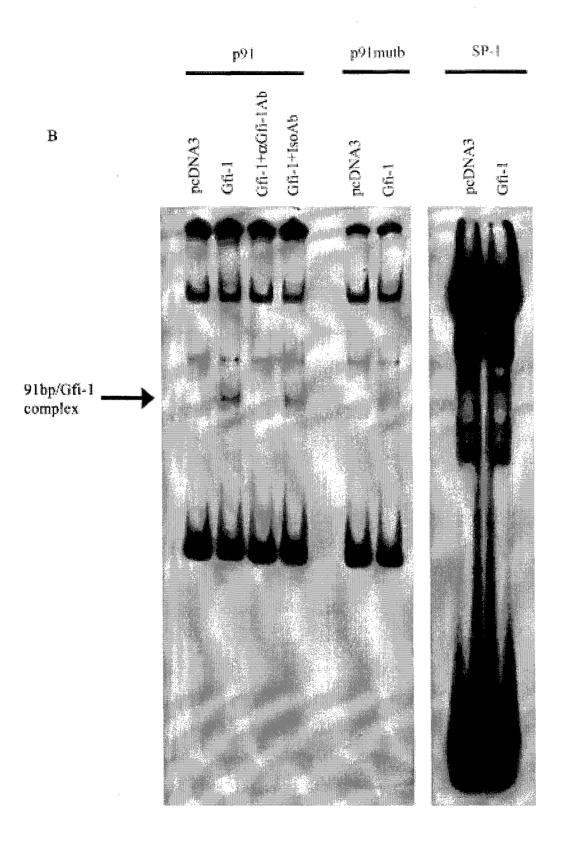
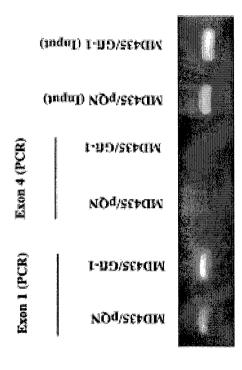


Figure 15B: Binding of Gfi-1 to exon 1 of GADD153. Experiments were done as in A. Gfi-1:DNA complex was detectable with nuclear extracts from cells transfected with gfi-1. This complex was disrupted by Gfi-1 specific antibody but not isotype control antibody. Also, binding of Gfi-1 to exon 1 with mutation of Gfi-1 binding site (p91mutb0 was substantially lower compared to p91 probe.



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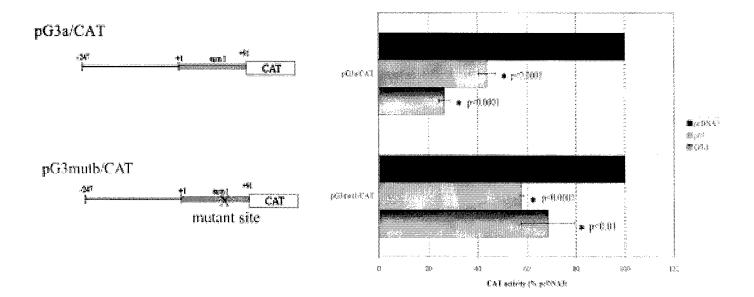


Figure 17: Gfi-1 represses GADD153 promoter. Cos-1 cells were transfected with pG3/CAT or pG3/CAT with mutation of Gfi-1 binding site (pG3mutb/CAT) and p65 or Gfi-1 expression vector. PCH110 was used as an internal control to measure transfection efficiency. pG3a/CAT but not pG3mutb/CAT activity was lower in cells transfected with p65 or Gfi-1. Thus, Gfi-1 functions as repressor of GADD153.

P59-14: NF-KAPPAR PROVIOTES ÉPÍTHÉLIAL-TO-MÉSENCHY-MAL TRANSITION OF IMMORTALIZED MAMMARY EPITHE-LIAL CELLS THROUGH ZEB-1 AND ZEB-2

Het Un Chus, Spell Bidve, and Haitkebâna Nakaberi Indian University School of Medicine, Indianopolis IN E-scall- Inst-hatfeborat ada

Epithelial to recombying transition (EMT) as a process for which apabelial colleacquire meanwhyreat properties, anothing these to migrate to other environment. EMT therefore plays as important role in embryogeneous as well as in the growth, asympton, and measurants of convey rolls. For example, EMT is observed in 18th of from concert and EMT has recomby been almost to generate meanwhymant atminewhich can support the growth of concercible. However, significant pathways that industry EMT in breast cancer is not completely understand.

The event shaller signal activated transcription factor NF-8B has recently been identified as a major player in a variety of cancers. Constitutive activation of NF-8B has been observed during the indig and hypothysis stages of becauteancer. To undergoing the role of NF-8B in the early stages of bress cases, the mannersy epithelial cell line MCF-10A overcapeliating a constitutionly active from of the pE5 schmitt of NF-8B was presented. The MCF-10A picts cells acquired a fibreblactic morphology, which is characteristic of cells andergoing EMT-Newton ANLF-10A cells coloured in a three-dimensional invests organized into polarized blocking incites executers, whereas MCF-10A, cells discribed as other studies. MCF-10A-pe5 cells formed discriming studies. MCF-10A-pe5 cells formed discriming studies. MCF-10A-pe5 cells formed an other studies. MCF-10A-pe5 cells discribed as other studies. MCF-10A-pe5 cells discribed as other studies. MCF-10A-pe5 cells discribed as chere to calculate the factor of the studies of the interesting accidance were also increased in MCF-10A-pe5 cells. In addition, MCF-10A-pe5 cells shared as increase a cellular modific cellular discriminal properties of the interestingly, the supercent studies of the interestingly of the interestingly, the supercent studies of the interestinal of E-radionin and induction of EMT, were showned in MCF-10A-pe5 cells. The induction of EMT, were showned in MCF-10A-pe5 cells. The induction of EMT, were showned in MCF-10A-pe5 cells. The induction of EMT, were showned in MCF-10A-pe5 cells. The induction of EMT, were showned in the repression of E-radionin and induction of EMT, were showned in MCF-10A-pe5 cells. The induction of EMT, and ZEB-2 and zeb-cutered in a breast cancer cell line following NF-8B activation. These passages and the NF-8B activation.

Original work apported by the U.S. Army Medical Research and Materiel Command with DAMD17-01-1-02/4

P59-15: THE ROLE OF CXCL12 AND CXCL14 CHEMOKINES IN EPITHELIAL STROMAL CELL INTERACTIONS BURING BREAST TUMOR PROCRESSION

Takarki Imamura, Missa Allinen, Min Hu, Andrea Richardean, Stuart Schrött, and Koenchi Polenk

Dans-Farber Correr faultute and Hervisch Medical School, Boston, MA E-mail: Rometia Polymbiodic faururet intu

Despite sampelling cell biological ancides and histografic logical observations greens. acting aromal cells in historing receive our knowledge of the genes that mediate strongs Changes and interactions among various cell types in breast charge, and then role in turner indication and progression is limited. Similarly the occurrence and rate of greater changes in streamal cells are varietiment. In order to gain issight into these querience, we sauly252 the comprehensive gene expression profiles of each cell type compresing normal breast rises said in aim and invasive breast continemas using Serial Analysis of Gene Expressive. Based on those data we described that extensive gene supressive charges occur to all cell types during cancer progression and a significant fraction of extend gener enough secreted proteins and acception. Despite the dramatic gener expension changes in all cell types, greatic attentions were only discretely in concerenitheliat cells The CXCL14 and CXCL12 observables systemposesed in terror epithelist refl. The CXULIA and CALLIA community symmetries in minimum myospithelial cells and appropriations, respectively, bind so receiptors on epithelial cells and induces ficin production, migration, and invasions. Thus chemitimes may play 1 role in became timeligeness by seeing as pessence factors. To test this hypothesis to the community of the community play a rote in crease authorizes on seasing or parameter moreover, are not tree inproved. So we generated destroyies of bosins concer and aspeculificial cell lines that averesponse these chemicalistics or destroyer and their especiation using alkNA. These cells were then analyzed in various in vitro and in vivo assays to determine the effect of chemaking on cellular growth, mustics, and amorismests. Our data support the hypothesis that the CXCL12 and CXCL14 chemokings act as postation thatian and influence because distinctinenesis, suggesting that the therapeutic targeting of these path ways may be a new approach for broad concer weatness.

The U.S. Army Medical Research and Material Commend analog WHILWHILL-L-QUED supported this need.

P59-16: ALTERED BREAST CELL PHENOTYPE, SIGNALI AND GENE EXPRESSION BY PRYSICAL PROPERTIES OF I EXTRACELLILAR MATRIX

Paulo É. Provenzoso and Paisis in J. Kenty Department of Francischery, University of Wisconne, Midison, Wi Local Interrocessive and a

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P59-17: STROMAL-EPITHELIAL INTERACTIONS, MAMMAI TISSUE ARCHITECTURE AND TUMOR ANGIOGENESIS

Enbriels I. Riccordiery, Januthen N. Laklus, and Valeric M. Wester Institute for 352525th and Engineering and Department of Pathologic University Principles (Ph. Lakehling, Ph. Lakehling

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American Association for Cancer Research 2003 Annual Meeting

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Hui Lin Chua, PhD (Refer to this abstract as # 100184) Indiana University School of Medicine 712 Barnhill Drive Indianapolis, IN 46202 USA

An NF-kB-repressible element in the 5-untranslated region of the pro-apoptotic GADD153/CHOP gene

Hui Lin Chua, Harikrishna Nakshatri, Indiana University School of Medicine, Indianapolis, IN.

Cells exposed to stresses such as nutrient deprivation, hypoxia, ultraviolet light and agents that damage DNA or disrupt endoplasmic reticulum function show induced expression of GADDI 53/CHOP, GADDI 53 is a member of the C/EBP family of transcription factors and promotes apoptosis following cellular stress. Interestingly, cellular stress also induces NF-kB. which promotes cell survival by inducing transcription of a number of anti-apoptotic proteins, such as Bel-2, Bel-XI., x-IAP and c-IAP-2. GADD153 mRNA expression was previously shown in this laboratory to be inhibited by the p65 transactivating subunit of NF-kB. Therefore, cells expressing constitutively active NF-kB, as is the case in 30% of breast cancers, may be predisposed to transformation or malignancy, due to p65-mediated inhibition of GADD153 expression following cellular stress. By transient transfection assays, we identified an NF-kB-repressible element (NRE) within a 91-base pair portion of the 5-untranslated region of the GADD153 gene. In addition, transfection of p65 with various co-activators or corepressors fulled to reverse the inhibitory effect. Insertion of GADD153 NRE, but not an unrelated 91-base pair sequence, to the 5-untranslated sequence of the Rous Sarcoma Virus enhancer/promoter-CAT reporter (RSV/CAT) led to p65-dependent repression of RSV/CAT expression. Interestingly, the same region of GADD153 has been previously shown to be required for UVC-mediated repression of stress-inducible GADD153 expression. We did not observe direct binding of the p63 subunit or any p65-inducible proteins to the 91-base pair DNA sequence by electrophoretic mobility shift assay. Based on a previous observation of MyoD mRNA destabilization by p65, we speculate that a p65-inducible protein destabilizes GADD153 mRNA by binding directly to the 5-untranslated region of GADD153 mRNA. Understanding the mechanisms by which NF-kB mediates inhibition of GADD153 expression will facilitate the design of strategies to promote apoptosis of damaged cells thereby preventing initiation of transformation,

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Critent Problems in

Volume 26 Number 5 September/October 2002

NF-kB and Breast Cancer

Introduction	28.
NF-kB	28.
NF-kB Target Genes	28.
NF-kB in Mammary Gland Development	28(
Constitutive Activation of NF-KB in Breast Cancer	28.
Mechanisms of Constitutive NF-rcB Activation	286
NF-xB Target Genes in Breast Cancer	286
NF-kB in Breast Cancer Initiation, Progression,	29(
and Metastasis Initiation Progression Metastasis	29.29
NF-4R in Osteolytic Bone Lesions and Cachexia	299
NF-xB-Mediated Chemoresistance	367
Inhibitors of NF-xB as Chemosensitizers for	295
Breast Cancer Parthanolida	207
Antioxidants	767
IKK Inhibitors	296
Proteosome Inhibitors	297
Glucocorticoids	297
PPAR γ Agonists	297
Antisense Oligonucleotides	298
Potential Complications of Long-Term NF-kB Inhibition	38

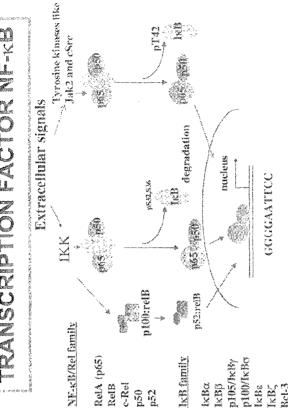
NF-KB and Breast Cancer

reast cancer is the leading cause of death among women in North America, with a 12% lifetime risk.1 Although there has been considerable progress in early detection and treatment, about 50% of patients eventually die of cancer.2 Death in most cases is due to metastasis and resistance to chemotherapy. Understanding the mechanism of metastasis and chemoresistance is critical for developing molecular arget-based therapies for breast cancer.

Recent developments in microarray technology have enabled molecular profiling of cancers. It is proposed that molecular profiling will offer more accurate predictions of future metastasis and treatment options,24 Mo-Gleevec, and Herceptin are some of the examples that are targeted against epidermal growth factor receptor (EGFR), Bcr-Abl, and the her2/Neu oncogene, respectively, 5-7 Unfortunately, most cancers undergo multiple changes, which makes it difficult to design molecular target-based herapy. An alternative approach to these cancers is to identify and target transcription factors that control the expression of genes involved in cancer initiation, progression, metastasis, and chemoresistance. In this article, we discuss the specific roles of nuclear factor-kB (NF-kB), a ranscription factor, in the regulation of genes involved in tumorigenesis ecular profiling also permits selection of targets for therapy. Iressa, and chemoresistance. We also highlight recent progress in the development of inhibitors of NF-kB as chemosensitizers for breast cancer.

ZF.KB

adhesion molecules, and cell cycle regulatory and antiapopiotic genes. NF-_KB was first identified in 1986 as the transcription factor involved in B lymphocyte-specific expression of k-immunoglobulin light chain.3 DNA binding activity was not detected in the nuclei of pre-B cells but could be activated by a posttranslational mechanism in these cells. Subsequent studies revealed ubiquitous activity of NF-κB and >180 target genes, including a variety of cytokines, cytokine receptors, cell Unlike general transcription factors, NF-kB activity in normal cells is



factors, cytokines, chemotherapeutic drugs, and oxidative stress activate IKK or a tyrosine kinase Fig 1. Three distinct mechanisms of NF-v3 activation: Extracellular signals, which include growth cascade leading to unmasking of nuclear localization signal and nuclear translocation of NF-xB.

ightly controlled, regulated primarily at the level of cytoplasmic and nuclear localization. A schematic view of this regulation is depicted in Fig

type-specific homodimeric and heterodimeric complexes of NF-κB have NF-kB is sequestered in the cytoplasm through association with a protein The NF-kB family consists of five members, each with a unique 500-amino acid-long dimerization domain called the rel homology (p105/p50), NF-xB2 (p100/p52), RelA (p65), RelB. and c-Rel. Although been identified.11 In rosting cells, with the exception of mature B cells. inhibitor-of- κB (IkB). There are six IkB proteins, which includes IkB α , IκBβ, IKBγ, IKBϵ. IKBϵ and Bel-3. ¹²⁻¹⁴ The highly conserved sequence called ankyrin repeats of InB mask the nuclear localization signal of domain. This domain is required for dimerization, nuclear translocation, and DNA binding.9 Members of the NF-kB family include NF-kB1 the p65;p50 heterodimer is the most predominant, several distinct cell NF-kB, thus retaining them in the cytoplasm. The p105 and p100

proteins, which are precursors for p50 and p52, respectively, have similar ankyrin repeats and function similar to LkB before processing. 12

unmasking of a nuclear localization signal of the p65:p50 heterodimer. $^{17.20}$ The second pathway involves phosphorylation of IkB α at described in several recent reviews. 15-20 Briefly, NF-kB is activated by a apeutic drugs (taxol, doxorubicin, camptothecin), 16,17 These inducers The mechanism of NF-kB activation has been studied extensively and number of stimuli, which includes cytokines (tumor necrosis factor [TNF] amily including receptor activator of NF-κB ligand [RANKL], interleucin [IL]-1, IL-17, IL-18), growth factors (EGF, PDGF, EPO, insulin, heregulin), physiologic mediators (angiotensin II, platelet activating reactive oxygen species (hydrogen peroxide), stress (UV irradiation, pH, hypoxia, heavy metals), apoptosis mediators (anti-Fas), and chemotheractivate at least three distinct pathways leading to activation of NF-rcB Fig 1). The majority of the inducers activate the IkB kinase complex (IKK), which phosphorylates $I\kappa B\alpha$ at serine residues 32 and 36. Phosshorylated $1\kappa B\alpha$ is degraded by the 26S proteosome leading to the tyrosine 42, leading to dissociation of the IkB α :NF-kB complex without cessing of p100 to p52 within the p100:relB complex, which results in NF-KB then moves into the nucleus and binds to response elements in the tion, and coactivator and corepressor interactions. 20,23,24 NF-kB termiion, heterodimerization among different NF-kB family members, and a factor), bacterial infection (Staphylococcus aureus), viral infection (HIV-1), bacterial and viral products (lipopolysaccharide, hemagglutinin), degradation of IkBa. 21 The third pathway involves enhanced processing of p100 upon phosphorylation by IKK α . Phosphorylation triggers prounmasking of nuclear localization signal of p52:relB heterodimer.22 promoter region of its target genes and activates transcription. NF-kB activity in the nucleus is further regulated by phosphorylation, acetylanates expression of its target genes by inducing IkBa expression, and the newly synthesized IkB α sequesters NF-kB back into the cytoplasm.²⁵ Thus, several regulatory steps including controlled subcellular localizasion by NF-kB. Any aberration in these steps can lead to uncontrolled gene expression by NF-kB, with profound impact on cell growth, negative feedback loop control tissue-specific and inducible gene expreslifferentiation, apoptosis, and immune response.

NF-kB Target Genes

NE- κ B regulates expression of genes involved in diverse functions; a partial list of genes that have potential impact on cancer is shown in Table 1. These genes control cell proliteration (cyclin D1, c-Myc), cell adhesion

TABLE 1. NF-x8--Regulated genes relevant for cancer

	Function
NF-xB-inducible genes	
Cyclin D1	Proliferation
c-Myc	Proliferation
ICAMI	Cell adhesion
VCAM-1	Cell adhesion
ELAM-1	Cell adhesion
Integrin aV	Cell adhesion, proliferation, metastasis
TNFa	Angiogenesis, cachexia, osteoclast activation
L-1	Angiogenesis, cachexía, osteoclast activation
E-6	Migration, angiogenesis, metastasis,
\$;	Cacrexia
œ-l	Angiogenesis, cachexía
iNos	Proliferation, angiogenesis, metastasis
COX-2	Proliferation, angiogenesis, metastasis
MIP-1, MIP-2	Migration, anglogenesis, osteoclast activation
M-CSF	Osteoclast activation
VEGF	Angiogenesis
Gro-1	Metastasis, angiogenesis
UPA	Invasion, migration, metastasis
MMP9	Metastasis
Haparanase	Metastasis
Bct2	Antiapoptosis, chemoresistance
BCI-XL	Antiapoptosis, chemoresistance
A1/841	Antiapoptosis
XIAP	Antiapoptosis
clAP-1, clAP-2	Antiapoptosis
TRAFL, IRAF2	Antiapoptosis
CFLIP	Antiapoptosis
Mn-SOD	Antiapoptosis, chemoresistance
Glutathione synthase	Antiapoptosis, chemoresistance
GADD458	Antiapoptosis
AKT1	Antiapoptosis, chemoresistance
NF-kB-repressible genes	
GADD153/CHOP	Proapoptosis
TIMP1, TIMP2	Metastasis suppressor
Plasminogen activator inhibitor	Metastasis suppressor

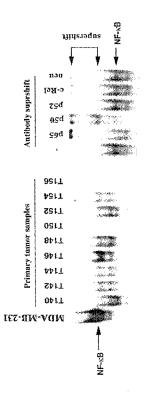
(ICAM-1, VCAM-1, ELAM1, Integrin αV), immume response (TNFα, IL-1α, IL-1β, IL-2, IL-6, IL-8, chemokines such a MIP1α, MCP-1), cell migration (urokinase plasminogen activator [uPΛ], matrix metalloproteinase 9), angiogenesis (VEGF) and antiapoptosis (Bcl-2, Bcl-X_L, A1/Bfl1, A20, xIAP, clAP-1, cIAP-2, TRAF-1, TRAF-2, cFLIP, manganese superoxide dismutase, γ -glutathione synthase, and GADD45β). $^{16,26-40}$ Several of these genes are involved in the normal developmental processes. Consistent with the role of NF-κB in embryonic development, "knocking-out" of

the *RelA* subunit of NF-**kB** leads to embryonic lethality.⁴¹ Similar defects were also observed in mice lacking IKK activity.^{42,43} Knockout mice lacking NF-**kB1**, NF-**kB2**, and c-*Rel* develop normally but show defects in the immune response.^{44,47} Osteopetrosis develops in mice lacking both NF-**kB1** and NF-**kB2**, suggesting a role for NF-**kB**-regulated genes in osteoclast activation.⁴⁸

NF-kB in Mammary Gland Development

lactation, and reactivated during involution. 51,52 However, reactivation of ing IkB α -deficient mammary epithelium into the mammary fat pad of Jifferentiation during lactation, and extensive apoptosis of the secretory epithelial cells during involution. 49.50 Mouse and rat models have been used to determine the potential role of NF-kB in these processes. NF-kB NF-kB during involution occurs exclusively in nonapoptotic epithelial cells, which suggests a prosurvival function of NF-kB during involumammary gland remains controversial, as the p65:p50 heterodimer and the p50 homodimer have both been reported as activated by different groups. 53,54 Nonetheless, the importance of NF-kB in proliferation and wild-type mice. 55 IkBlpha-deficient cells, in which NF-kB is constitutively localized in the nucleus, showed increased proliferation, lateral ductal type counterparts. Furthermore, the extracellular matrix was reduced adjacent to $I\kappa B\alpha$ -deficient epithelium. Thus, it appears that NF- κB is required for proliferation of mammary epithelial cells, blocking of Development of the mammary gland is a complex process involving DNA binding activity is elevated during pregnancy, suppressed during tion.⁵³ The identity of subunits of NF-κB activated in the normal branching of mammary epithelial cells has been confirmed by transplantbranching, and pervasive intraductal hyperplasia compared with wildextensive proliferation of mammary epithelial cells during pregnancy, terminal differentiation, and/or delaying the onset of apoptosis.

TNF α and RANKL are the major physiologic activators of NF- κ B in the mammary gland. ^{56,57} A systematic genetic analysis in mice revealed that activation of NF- κ B and the resulting proliferation of mammary epithelial cells during pregnancy occurs through a linear cascade: RANKL \rightarrow RANK \rightarrow IKR $\alpha\rightarrow$ IKB $\alpha\rightarrow$ NF- κ B \rightarrow Cyclin D1. ⁵⁷ Deletion or functional inactivation of RANKL, RANK, IKK α , or cyclin D1 in mice results in similar defects in proliferation of mammary epithelial cells. With respect to tumorigenesis, any genetic mutation that leads to aberrant activation of this cascade could lead to increased proliferation and predispose mammary epithelial cells for transformation.



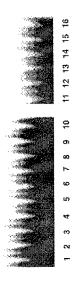


Fig. 2. NF-4B DNA binding activity in primary breast cancer. NF-4B DNA binding activity was measured by electrophoretic mobility shift assay. MDA-MB-231 cell extract was used as positive control. NF-4B DNA complex contains p65 and p50 subunits as determined by antibody supershift assay [lanes 11 to 16].

Constitutive Activation of NF-kB in Breast Cancer

Progression of breast cancer from a benign to a malignant phenotype is accompanied by overexpression of several growth factors, cytokines, and chemokines by cancer cells. Some of these growth factors and cytokines can induce the expression of prometastatic genes through NF- κ B. One example is uPA, whose overexpression correlates with poor clinical outcome, ⁶¹ These observations prompted our laboratory to investigate whether NF- κ B is constitutively active in breast cancer. NF- κ B DNA binding activity was examined in a panel of established breast cancer cell lines. In general, elevated NF- κ B DNA binding activity was observed in breast cancer cell lines with invasive and metastatic growth properties. ⁶² Furthermore, extracts from primary breast tumors showed elevated NF- κ B DNA binding activity (Fig 2). Increased expression of IL-6 and uPA was observed in breast cancer cell lines with constitutive NF- κ B DNA binding activity (Fig 2). ⁶³ Sovak et

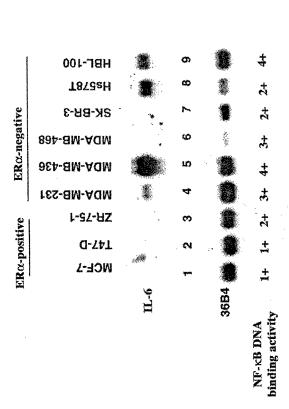


Fig. 3. Breast cancer cells with constitutive NFxB overexpress urokinase plasminogen activator. Urokinase plasminogen activator expression was measured by Northern blotting. Ribosomal protein gene 36B4 was used as control.

al⁶⁴ reported constitutive NF-κB DNA binding activity in 65% of primary breast cancers. The NF-κB:DNA complex that we observed in cell lines as well as primary breast cancers was predominantly a heterodimer of p65:p50 (Fig 2). Constitutive nuclear localization of p50, p52, c-*Rel*, and *Bcl*-3 and overexpression of p100/p52 in breast cancer have been reported by others. ^{65,66}

It was apparent in our studies that the majority of breast cancer cells with constitutive NF- κ B DNA binding and transcriptional activity are estrogen receptor- α (ER α)-negative.⁶² Because the ER α -negative phenotype correlates with poor prognosis, we explored the possibility that ER α negatively regulates NF- κ B activity in breast cancer cells. Indeed, although DNA binding of NF- κ B is inducible in ER α -positive breast cancer cells, NF- κ B is transcriptionally less competent in these cells.⁶² ER α physically associates with DNA-bound NF- κ B and interferes with its transactivation.⁶⁷ This function of ER α , which is called transrepression, is essential to prevent invasion by cancer cells.⁶⁸ This may explain the differences in the metastatic potential of ER α -positive and ER α -negative breast cancers. Approximately NF- κ B is more relevant to ER α -negative breast cancers. Approximately

30% of breast cancers are ER α -negative at the time of initial diagnosis, and the 5-year survival rate for these patients is <50%.⁴ We propose NF- κ B as the molecular target for ER α -negative breast cancers.

Mechanisms of Constitutive NF-kB Activation

in both cancer and stromal cells. Neutralizing antibodies against LL-1 α cell lines derived from transgenic mice that specifically overexpress aeregulin in constitutive activation of NF-kB in breast cancer cells. 69-73 Biswas et al72 showed a correlation between EGFR overexpression and increased NF-kB activity in ERα-negative breast cancers. Inhibitors of blocked NF-kB activity and reduced NF-kB-regulated gene expression in both cancer and stromal cells, 73 To identify the specific oncogenes that may be involved in NF-kB activation, we recently used mammary fumor oncogenes in their mammary gland. Constitutive NF-kB DNA binding from c-Myc, ras, or het2/neu transgenic mice. 70 However, her2/neu potentiated TNF-induced NF-kB activation and stimulated cell growth in the presence of TNF. Interestingly, overexpression of EGFR, heregulin, and her2/neu is usually observed in $ER\alpha$ -negative breast cancers and is associated with poor prognosis. 74,75 Thus, it is likely that NF-κB is the downstream target of signal transduction pathways that are responsible We and others have shown the involvement of EGFR, IL-1 α , and EGFR reduced NF-κB activity and cell growth. We observed that a subset of ER α -negative breast cancer cells secrete IL- 1α , which activates NF- κB activity was observed in cell lines from heregulin transgenic mice but not for growth of ERa-negative breast cancers.

Apart from EGFR. IL-1a, heregulin, and her2/neu, breast cancers express bone morphogenic factor 4, PDGF. and transforming growth factor-a. ⁷⁶⁻⁷⁹ These growth factors may be involved in autocrine and paracrine induction of NF-κB in the tumor microenvironment. However, their contribution in constitutive NF-κB activation in breast cancer is yet to be verified. In addition to production of growth factors and cytokines, hypoxic and acidic pH conditions are common in the tumor microenvironment, and both of these conditions may contribute to constitutive NF-κB activation in breast cancer. ⁸⁰ Furthermore, elevated expression and/or activity of IKK and casein kinase II is linked to constitutive NF-κB activation in breast cancer. ⁸¹

NF-kB Target Genes in Breast Cancer

Although more than 180 genes are induced by NF- κ B, not all of them are expressed in breast cancer. To identify NF- κ B-regulated genes in breast cancer cells, we created MDA-MB-231 breast cancer cells over-

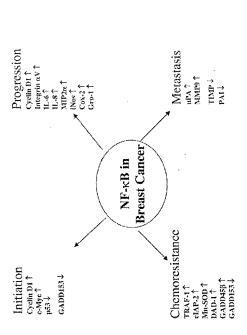


Fig. 4. Potential contributions of NF-κB in breast cancer initiation, progression, metastasis, and chemoresistance, NF-κB-regulated genes involved in breast cancer are shown.

expressing $I\kappa B\alpha$. 82 MDA-MB-231 cells contain constitutively active NF- κB because of IL-1 α expression and EGFR overexpression. $^{62.69}$ Comparative analysis of parental and $I\kappa B\alpha$ -overexpressing cells by regulated genes. All but one of them are induced by NF-kB. Genes induced by NF-kB include uPA, IL-6, IL-8, macrophage inhibitory protein-2- α (MIP2 α), defender against cell death (DAD), ICAM-1, leinase (MMP)9, VEGF, cyclin D1, and Gro-1.3.4.83.84 We have also glutathione S-transferase M4, Cu/Zn superoxide dismutase 1, c-IAP2, TRAF-1, and Mn-SOD, 63.82 NF-kB-inducible genes in breast cancer identified by microarray analysis by others include matrix metalloproobserved repression of GADD153/CHOP by NF-kB in breast cancer cells. 85 We expect further additions to the list of genes induced by NF-κB in the near future. The potential role of these genes in breast cancer cDNA microarray and RNase protection assay identified several NF-kBnitiation, progression, metastasis, and chemoresistance is shown diagrammatically in Fig 4.

NF-kB in Breast Cancer Initiation, Progression, and Metastasis

Initiation

DMBA- or benzo[a]pyrene-induced mammary tumors in rats and in vitro transformation assay of human mammary epithelial cells obtained

Curr Probl Cancer, September/October 2002

ion during initiation of breast cancer.86 With both assays, activation of cancers, appears to be the target of NF-κB in this system.86 NF-κB may ion. 26.27.87 Cyclin D1 is overexpressed at the early stages of breast cancer from reduction mammoplasty have been used to analyze NF-kB activathe p50:p65 heterodimer was observed before malignant transformaion. 86 The c-Myc oncogene, which is overexpressed in 30% of breast lirectly increase cyclin D1 expression, whereas c-Myc may increase the expression of cyclin A and cyclin E, leading to rapid cell proliferaprogression, such as ductal carcinoma in situ but not in premalignant esions (ie, LCFS). 88.89 Cyclin D1 overexpression in ERα-negative but not ERa-positive breast cancers correlates with poor prognosis.90 Furhermore, cyclin D1 is absolutely necessary for her2/neu and ras oncogene-induced manmary tumor formation, 91 Thus, growth factor/ oncogenic pathways activated early in premalignant cells (particularly ERα-negative) may utilize NF-κB to induce cyclin D1 expression and ransformation.

Modulation of p53 activity may be another mechanism by which NF-kB promotes breast cancer initiation, p53 is a tumor suppressor gene that unctions as a gatekeeper by arresting cells with damaged DNA at the G0/G1 phase of the cell cycle. If DNA damage is less extensive, it is repaired and cells progress through the cell cycle. If DNA damage is extensive, p53 triggers apoptosis. ⁹² Several recent publications, with one exception, indicate that NF-kB inhibits p53 activity. 93-95 Thus, by reducing p53 activity, NF-κB may enhance survival of cells with damaged DNA. Surviving cells with damaged DNA accumulate additional mutations, leading to transformation. Our recent studies indicate that NF-kB represses expression of the GADD153/CHOP gene.85 GADD153/CHOP is usually induced in cells exposed to environmental toxins and during stress of endoplasmic reticulum seen with several diseases.96 Depending on the severity of DNA damage and stress. GADD153 triggers apoptosis. By reducing GADD153 expression, NF-κB may promote survival and transformation of cells with damaged DNA.

Progression

Progression of cancer requires proliferation, angiogenesis, and protection against apoptotic signals originating from the immune system. Several of the NF-κB-regulated genes can aid in this process. For example, NF-κB can increase cell proliferation through c-*Myc* and cyclin D1. NF-κB can promote angiogenesis by inducing the expression of IL-6, IL-8, *Gro*-1, and VEGF ^{9,16,97,99} NF-κB also affects angiogenesis indirectly through stromal cells. We have observed that IL-1α secreted by

breast cancer cells induces NF-κB in stromal fibroblasts, resulting in increased expression of IL-6 and IL-8 by these cells.^{69,73} IL-6. although not directly linked to angiogenesis, can induce the expression of tissue factor 1 in stromal cells.¹⁰⁰ Tissue factor 1 is a potent inducer of angiogenesis.¹⁰¹ Finally, NF-κB-regulated genes, including Mn-SOD, cIAP-2, DAD-1, and TRAF-1, can protect cancer cells from the cytotoxic effects of TNF.^{35,102,103} It was reported recently that normal mammary epithelial cells secrete Fas ligand, which triggers apoptosis of surrounding cancer cells.¹⁰⁴ However, cancer cells can protect themselves against Fas ligand through activation of NF-κB.¹⁰⁴

The serine/threonine kinase AKT functions as a major survival protein for a variety of cell types. ¹⁰⁵ Overexpression of this kinase alone is sufficient to induce mammary gland hyperplasia. ¹⁰⁶ It was demonstrated recently that AKT is one of the target genes of NF-κB. ¹⁰⁷ Thus, constitutive NF-κB activation may lead to elevated AKT, resulting in prolonged survival of immortalized and/or transformed cells in the mammary gland. Consistent with this possibility, AKT is overexpressed in breast cancer.

Additional candidate NF-κB-regulated genes involved in breast cancer progression include inducible nitric oxide synthase and COX-2, both of which are overexpressed in advanced breast cancers. ^{110,111} Overexpression of COX-2 and inducible nitric oxide synthase correlates with increased angiogenesis and metastatic disease. ^{110,112} COX-2 overexpression, which is observed in ~40% of breast cancers, is associated with the ERα-negative phenotype and unfavorable prognosis. ¹¹³

Metastasis

The metastasis of malignant cells is a multistep process involving (1) extracellular matrix degradation and subsequent detachment of cancer cells from the primary tumor; (2) invasion of surrounding tissue; (3) penetration of blood and/or lymphatic vessels (also called intravasation); (4) survival in the blood circulation, transport to new sites, and arrest of cancer cells in the microcirculation; (5) migration of cancer cells through the vessel wall into the interstitial space (extravasation); and (6) proliferation of cancer cells in the target organs (colonization). Two mutually dependent protease systems are required for extracellular matrix degradation. The first system includes MMP-1, MMP-2, MMP-3, MMP-9, and MMP-13. These proteins are secreted as pro-MMPs, and most of them are activated by the second protease system. The second system involves the uPA/urokinase plasminogen activator receptor (uPAR)/plasminogen system. The uPA/uPAR converts plasminogen to

plasmin, which activates MMPs. ¹¹⁵ The uPA/uPAR and MMP-9 are essential for intravasation of a number of cancer cell lines, including MDA-MB-231 breast cancer cells. ¹¹⁶ Although overexpression of uPA/ uPAR/MMP-9 is not necessary for survival and attachment of cancer cells to the microvasculature, overexpression of uPA/uPAR is essential for migration and colonization at the target site. ¹¹⁶⁻¹¹⁸ NF-κB can promote extracellular matrix degradation, migration, and colonization at the target site by directly inducing the expression of uPA and MMP-9, thereby increasing metastasis. ^{33,34,119} NF-κB can also increase MMP-2 indirectly by synergizing with the AP-1 transcription factor. ⁶⁷ NF-κB has also been shown to increase the expression of prometastatic heparanase and decrease the expression of antimetastatic tissue inhibitors of MMP-1 and MMP-2 and plasminogen activator inhibitor 2. ¹²⁰ Consistent with the role of NF-κB in the degradation of extracellular matrix, mammary glands of mice transplanted with IκBα-deficient mammary epithelial cells showed a lower level of intact extracellular matrix. ⁵⁵

NF-KB in Osteolytic Bone Lesions and Cachexia

While cachexia contributes to mortality in patients, osteolytic lesions do not. Bone metastasis develops in ~70% of patients with breast cancer, whereas cachexia develops in 30% of patients. ^{121,122} Several factors, including IL-1, IL-6, RANKL, parathyroid hormone–related protein (PTHrP), MIP1α, and macrophage colony stimulating factor (M-CSF), have been implicated in osteoclast activation. ^{123,124} Cancer cells secrete most of these factors, and the expression of some of these factors (IL-1, IL-6, MIP1α, and M-CSF) is dependent on NF-κB. In fact. MDA-MB-231 breast cancer cells, which have been used extensively for studying osteolytic bone metastasis in a xenograft model, express IL-1. IL-6, PTHrP, and M-CSF. ^{125,126} We have observed that expression of IL-1 and IL-6 in these cells is dependent on NF-κB. Thus, extensive bone loss can be anticipated when breast cancer cells, with constitutively active NF-κB, increase the level of osteoclastogenic factors in the bone microenvironment after metastasis.

IL-6, IL-8, TNF α , interferon- γ (IFN γ), leukemia inhibitory factor. myostatin/GDF8, protein mobilizing factor (PMF), and lipid mobilizing factor (LMF) are some of the factors involved in cachexia. ¹²⁷⁻¹²⁹ IFN γ , in combination with TNF α , has been shown to induce cachexia through NF- κ B-dependent destabilization of MyoD mRNA in muscle. ¹³⁰ Because expression of IL-6, IL-8, TNF α , and IFN γ is dependent on NF- κ B, it is possible that breast cancers with constitutively active NF- κ B overexpress these cachexia-inducing factors.

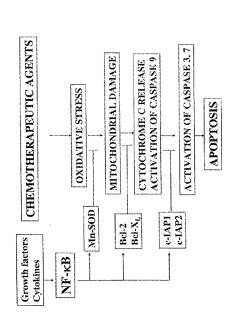


Fig 5. Model depicting mechanism of chemoresistance by NF-xB.

NF-kB-Mediated Chemoresistance

It was recognized a decade ago that NF-kB is activated in cancer cells daunorubicin, and camptothecin. 131-133 These initial results provided a link between cell death and NF-kB activation. However, a definitive role repressor (a mutant form of IkBlpha in which S32 and S36 are mutated to nonphosphorylatable alanine and hence nondegradable), were found to be more sensitive to daunorubicin and TNF. ¹³⁴⁻¹³⁶ Subsequent studies have shown that NF-kB is essential for the expression of several antiapoptotic genes including Bcl-2, $Bcl-X_L$, TRAF-1, TRAF-2, Bfl1/A1, cFlip. XIAP, cIAP-1, cIAP-2, and $Mn-SOD^{-16,17,35}$ A schematic view of how these Chemotherapeutic drugs usually cause oxidative stress, leading to mitocaspase 9, releasing active caspase 9,137-139 Caspase 9 then cleaves procaspase 3 to caspase 3, which is an executioner caspase. Caspase 3 Caspase 3 cleaves several important proteins, including PARP. Mn-SOD overexpression of Mn-SOD alone in breast cancer cells is sufficient for was not established until 1996. Cells lacking NF-kB activity, obtained either through gene knockout or through overexpression of $I\kappa Blpha$ superproteins may confer resistance to chemotherapy is shown in Fig 5. chondrial damage. Cytochrome C is released from damaged mitochondria, which activates the apoptosome. The apoptosome cleaves proactivates specific DNases involved in cleavage of chromosomal DNA. after exposure to chemotherapeutic agents including taxol. doxorubicin, confers resistance to chemotherapy by reducing oxidative stress. Indeed, for NF-kB in cell survival, with or without exposure to chemotherapy.

death by preventing activation of caspase 9 as well as by inhibiting the activity of caspase 3.138 Thus, NF- κ B-regulated genes confer resistance to chemotherapy by interfering at multiple sites in the normal pathway of resistance against taxol, H₂O₂, and okadaic acid. ¹⁰² Bcl-2 and Bcl-X_L confer resistance to chemotherapy by preventing the release of cytochrome C from the mitochondria. XIAP, cIAP-1, and cIAP-2 reduce cell

of antiapoptotic genes TRAF-1, cIAP-2, DAD-1, and Mn-SOD was lower in $I_K B \alpha S R$ cells compared with parental cells. We then examined the sensitivity of parental and IkB α SR cells to taxol. IkB α SR cells were more sensitive to taxol-induced G2/M arrest and apoptosis.⁸² Thus, NF-kB-regulated genes appear to control the response of breast cancer To understand the role of NF- κ B in resistance to chemotherapy, we generated MDA-MB-231 cells overexpressing I κ B α SR. ⁸² The expression cells to taxol.

gesting the requirement of NF-kB for chemotherapy-induced death of sistance. In fact, adenovirus-mediated gene therapy with $I\kappa B\alpha SR$ has in a xenograft model. 140 To avoid the difficulty in targeting $IkB\alpha SR$ to inhibitor derived from the herb feverfew, 141 increased the sensitivity of MDA-MB-231 and HBL-100 cells to taxol.82 Weldon et al¹⁴² made 11-7082 increased sensitivity of MCF-7 APO+ breast cancer cells to been shown to increase sensitivity of colorectal carcinoma cells to CPT11 inhibitors of NF-kB as chemosensitizers to taxol. Parthenolide, an NF-kB similar observations. Pharmacologic inhibition of NF-kB with BAY doxorubicin and taxol. Although there is one contradicting report sugpreast cancer cells, 143 most of the current literature favors the antiapopotic function of NF-κB in breast cancer and points to a new therapeutic Gene therapy with $I\kappa B\alpha SR$ is one mechanism of overcoming chemorebreast cancers, we evaluated the utility of various exogenous chemical direction.

Inhibitors of NF-kB as Chemosensitizers of **Breast Cancer**

anti-inflammatory action of glucocorticoids was due to inhibition of NF-κB activity. ^{144, 145} Subsequently, a number of natural and synthetic scribed in the literature. 15 These include folk medicines derived from natural sources, known for their anti-inflammatory properties, which have been in clinical use for many years. It was reported in 1995 that the compounds with anti-inflammatory properties have been tested for unti-NF-kB activity, and the anti-inflammatory medicinal plants from the More than 100 compounds with anti-NF-кВ activity have been de-

genera Barbetis, Coptis, and Gravel Root are receiving renewed attention. ^{146,147} Below, we describe some of the compounds, which may be explored as chemosensitizers for breast cancer.

Parthenolide

Parthenolide is the active ingredient of the herb feverfew. ¹⁴¹ Feverfew is being used as a migraine prophylaxis. ¹⁴⁸ It inhibits NF-κB activation by reducing IKK activity. ¹⁴⁹ Parthenolide also inhibits DNA binding of the STAT family of transcription factors, which probably is an indirect result of NF-κB inhibition. ¹⁵⁰ At a higher concentration, parthenolide inhibits MAP kinase activity. ¹⁵¹ After obtaining encouraging in vitro results, we are exploring the combination of parthenolide and taxanes in a xenograft model of breast cancer. In our nude mice studies, we did not observe toxic side effects of prolonged parthenolide treatment. One limitation of parthenolide is its relatively short half-life. It inhibited NF-κB DNA binding activity for only 5 hours. ⁸⁵ It may therefore be necessary to develop more stable analogues. Nonetheless, we remain optimistic that our ongoing studies will reveal a useful role for parthenolide as a chemosensitizer.

Antioxidants

Antioxidants with NF-kB-inhibitory properties include curcumin and epigallocatechin-3-gallate (EGCG). ^{152,153} Curcumin is a common ingredient in Asian cooking and has been tested in animal models as a chemoprevention agent. ^{154,155} EGCG is the active ingredient of green tea and is currently being tested as a chemoprevention agent also. ¹⁵³ Although these two compounds are promising, their use as chemosensitizers may be limited because relatively higher concentrations (30 to 60 µmol/L) are necessary to achieve NF-kB inhibition.

IKK Inhibitors

Aspirin, ibuprofen, 4-hydroxy-2-nonenal, and sanguinarine are potent inhibitors of IKK. ¹⁵⁶⁻¹⁶⁰ It will be interesting to see whether prior treatment with aspirin or ibuprofen increases sensitivity of breast cancer cells to chemotherapy. Recently, a peptide inhibitor of IKK activity has been described and may function as a chemosensitizer. ¹⁵¹ The mechanism of NF-κB inhibition by several compounds is not known, and it is presumed that some of these inhibitors target IKK. IKK inhibitors hold more promise than other broad-spectrum inhibitors of NF-κB because of fewer undesirable side effects.

Proteosome Inhibitors

Inhibitors of the 26S proteosome (a cellular factory involved in controlled degradation of proteins), which block degradation of phosphorylated IkBa, have been used as NF-kB inhibitors for in vitro studies. These compounds include MG132, lactacystine. ALLnL, and cyclosporine A.¹⁵ PS341, which is in phase II and phase III clinical trials, is a potent inhibitor of NF-kB.¹⁶² The major molecular target of PS341 in melanoma is NF-kB. as constitutive activation of NF-kB is frequently observed in this cancer.¹⁶³ We believe that PS341 will have similar effects in breast cancer either as a single agent or in combination with chemotherapy.

Glucocorticoids

The anti-inflammatory activity of glucocorticoids is largely due to inhibition of NF-kB. ^{144,145} Inhibition of NF-kB activity by glucocorticoids involves two independent cell type–specific mechanisms. In certain cell types, glucocorticoids directly induce the expression of IkBa. thereby reducing nuclear translocation and DNA binding of NF-kB. ^{144,145} In most cell types, however, glucocorticoids inhibit NF-kB activity through transcriptional interference. ^{164,165} The glucocorticoid receptor, upon binding of glucocorticoids, translocates to the nucleus, where it interacts with the p65 subunit of NF-kB. The p65:glucocorticoid receptor complex is transcriptionally inactive. Elegant knock-in mutation studies have proved that this function of the glucocorticoid receptor is important for repression of the inflammatory response. ¹⁶⁶ Some of the chemotherapy combinations include glucocorticoids mainly to overcome treatment side effects. ^{167,168} It is possible that glucocorticoids enhance sensitivity of cancer cells to chemotherapy by reducing NF-kB-dependent expression of antiapoptotic genes.

PPARy Agonists

Peroxisome proliferator activator receptor-γ (PPARγ) agonists such as troglitazone are potent antidiabetic drugs in clinical use. ¹⁶⁹ PPARγ agonists are also currently being tested as chemoprevention agents. ^{170,171} These agonists can inhibit NF-κB activity by at least two mechanisms. PPARγ directly interacts with the p65 subunit of NF-κB and inhibits its transactivation function. ^{172,173} PPARγ agonists also induce the expression of the tumor suppressor gene PTEN. ¹⁷⁴ PTEN inhibits the activity of AKT, a kinase that increases NF-κB DNA binding as well as transactivation. ^{175,176} Because PPARγ is expressed in breast cancer cells, its agonists may function as chemosensitizers in breast cancer by reducing

the expression of NF- κ B-regulated genes. ¹⁷⁷ Consistent with this possibility, the natural PPAR γ agonist 15-deoxy- $\delta^{13.15}$ -prostaglandin J2 inhibits expression of the NF- κ B-inducible genes MCP-1 and GADD45 β and increases expression of the NF- κ B-repressible gene CHOP/GADD153 in MDA-MB-231 cells. ¹⁷⁷

Antisense Oligonucleotides

Antisense oligonucleotides against p65 have been used in the mude mice model to evaluate the role of NF-κB in tumorigenecity and experimental colitis. The antisense oligonucleotides reduced in vitro growth and tumor formation by breast cancer cells, osteosarcoma, melanoma, and colon carcinoma cells. ¹⁷⁸ Similarly, antisense oligonucleotides also reduced experimental colitis. ¹⁷⁹ Recently developed RNA interference (RNAi) technology is more promising than the antisense technique for functional inactivation of a gene of interest. ¹⁸⁰ RNAi is a short, 21-base-long, double-stranded RNA molecule corresponding to a gene of interest, which through an unknown mechanism triggers degradation of the specific RNA. We anticipate that this technique will become a more powerful tool in preclinical and clinical studies for inactivation of NF-κB.

Potential Complications of Long-Term NF-kB Inhibition

The importance of NF-κB in the immune system led to initial concerns in developing NF-κB inhibitors as therapeutic agents. This concern was further strengthened by knockout studies showing an important role of NF-κB in liver organogenesis. He Embryonic death caused by massive hepatic apoptosis was observed in p65 knockout mice. On the basis of these results, hepatotoxicity was predicted with NF-κB inhibitors. However. p65 does not appear to be required for the survival of adult hepatocytes because conditional knockout of p65 in adult mice is not deleterious, except for a modest increase in bacterial infection. The common use of drugs such as aspirin and glucocorticoids, which inhibit NF-κB activity, has lessened concern over the therapeutic use of NF-κB inhibitors, but much work remains.

NF-kB in Anti-Estrogen Resistance

Approximately 50% of breast cancers are ERα-positive at the time of diagnosis. Although most patients respond to anti-estrogen therapy, in vitro studies have demonstrated an eventual resistance to therapy, ¹⁸² Dr Robert Clarke's group has recently used a xenograft model to study the mechanism of resistance to pure anti-estrogen, ICI182,780. ¹⁸³ Resistance to ICI182,780 was accompanied by increased NF-κB activity. These

ICI-resistant cells were growth-inhibited by parthenolide, suggesting the requirement of constitutive NF- κ B for the survival of these cells. The mechanisms for constitutive NF- κ B activation in these cells remains to be determined. It is possible that the ER α lost its ability to transrepress NF- κ B activity because of posttranslation modification or altered stability.

Summary and Future Directions

30% of patients with breast cancer. Because NF-kB is downstream of a technology. Herceptin was the first successful molecular target-based ways, particularly in ERα-negative breast cancers, it is an ideal target for intervention. In this article, we have highlighted how NF-kB controls therapy. A number of NF-kB inhibitors derived from natural products have already been characterized. These NF-kB inhibitors provide a perfect opportunity for the rational design of clinical trials that combine NF-κB at nanomolar concentrations, compounds that inhibit NF-κB for a tion of the human genome project and the development of microarray therapy developed for breast cancer. Unfortunately, it is beneficial in only number of growth factor/oncogene-activated signal transduction pathbreast cancer initiation, progression, metastasis, and resistance to chemolonger duration, production of orally active compounds, and determining the ideal sequence and duration of administration. We believe that these Molecular target-based therapy is becoming a reality with the complealternative medicine with conventional medicine. However, several challenges still exist, including the identification of compounds that inhibit challenges will be overcome in the near future as both academia and industry devote considerable resources in basic and translational research nvolving NF-kB.

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REFERENCES

- Benshushan A, Brzeziuski A. Hormonal manipulations and breast cancer. Obstet
- Caldas C, Aparicio SA. The molecular outlook. Nature 2002;415:484-485.
- van 't Veer LJ, et al. Gene expression profiling predicts clinical outcome of breast cancer. Nature 2002;415:530-536. ci m
- Sorlie T, et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci U S A 2001;98:10869-4
 - Ranson M. ZD1839 (Iressa): for more than just non-small cell lung cancer. Oncologist 2002;7:16-24. wi.
- Ligibel JA, Winer EP. Trastuzumab/chemotherapy combinations in metastatic breast cancer. Semin Oncol 2002;29:38-43. Ć,
- Capdeville R. Buchdunger E, Zimmermann J, et al. Glívec (STI571, imatínib), a rationally developed, targeted anticancer drug. Nat Rev Drug Discov 2002;1:493r.:
- Sen R, Baltimore D. Inducibility of kappa immunoglobulin enhancer-binding protein Nf-kappa B by a posttranslational mechanism. Cell 1986;47:921-928. ∞
- Baeuerle PA. Henkel T. Function and activation of NF-kappa B in the immune system. Annu Rev Immunol 1994;12:141-179. 6
 - Ghosh S. May MJ, Kopp EB. NF-kappa B and Rel proteins: evolutionarily conserved mediators of immune responses. Annu Rev Immunol 1998;16:225-260. Ö
- Lembecher T, Muller U. Wirth T. Distinct NF-kappa B/Rel transcription factors are responsible for tissue-specific and inducible gene activation. Nature 1993;365:767-
- Baeuerle PA, Baltimore D. NF-kappa B: ten years after. Cell 1996;87:13-20.
- Whiteside ST, Epinat JC, Ricc NR, et al. I kappa B epsilon, a novel member of the I kappa B family, controls RelA and cRel NF-kappa B activity. EMBO J
- Yamazaki S, Muta T, Takeshige K. A novel IkappaB protein. IkappaB-zeta, induced by proinflammatory stimuli, negatively regulates nuclear factor-kappaB in the nuclei. J Biol Chem 2001;276:27657-27662. 4
- Epinat JC, Gilmore TD. Diverse agents act at multiple levels to inhibit the Rel/NF-kappaB signal transduction pathway. Oncogene 1999:18:6896-6909, 3
- Pahl HL. Activators and target genes of Rel/NF-kappaB transcription factors. Oncogene 1999;18:6853-6866. <u>.</u>
 - Garg A, Aggarwal BB, Nuclear transcription factor-kappaB as a target for cancer drug development. Leukemia 2002;16:1053-1068. 7
 - Karin M, Cao Y, Greten FR, et al. NF-kappaB in cancer: from innocent bystander to major culprit. Nature Rev Cancer 2002;2:301-310. 8
- Karin M. Lin A. NF-kappaB at the crossroads of life and death. Nat Immunol 2002;3:221-227. <u>5</u>
 - Ghosh S, Karin M. Missing pieces in the NF-kappaB puzzle. Cell 2002;109:S81-8

Surr Probl Cancer, September/October 2002

- Digicaylioglu M, Lipton SA. Erythropoietin-mediated neuroprotection involves cross-talk between Jak2 and NF-kappaB signalling cascades. Nature 2001;412:641-21.
- Senftleben U, et al. Activation by IKRalpha of a second, evolutionary conserved, NF-kappa B signaling pathway. Science 2001;293:1495-1499, 5
 - Baek SH, et al. Exchange of N-CoR corepressor and Tip60 coactivator complexes links gene expression by NF-kappaB and beta-amyloid precursor protein. Cell 2002;110:55-67. ä
 - Chen L, Fischle W. Verdin E. et al. Duration of nuclear NF-kappaB action regulated by reversible acetylation. Science 2001;293;1653-1657. ঠ
 - Haskill S, et al. Characterization of an immediate-early gene induced in adherent monocytes that encodes I kappa B-like activity. Cell 1991;65:1281-1289. 23.
 - Hinz M. et al. NF-kB function in growth control: regulation of cyclin D1 expression and G₀/G₁-to-S-phase transition. Mol Cell Biol 1999;19:2690-2698. 26.
- Guttridge DC, Albanese C. Reuther JY, et al. NF-kappaB controls cell growth and differentiation through transcriptional regulation of cyclin DI. Mol Cell Biol 1999;19:5785-5799. 27.
- Duyao MP, Buckler AJ, Sonenshein GE. Interaction of an NF-kappa B-like factor with a site upstream of the e- myc promoter. Proc Natl Acad Sci U S A 1990;87:4727-4731. 28
- van de Stolpe A, et al. 12-O-tetradecanoylphorbol-13-acetate- and tumor necrosis factor alpha-mediated induction of intercellular adhesion molecule-1 is inhibited by dexamethasone; functional analysis of the human intercellular adhesion molecular-1 promoter, J Biol Chem 1994;269:6185-6192. 5
- Jademarco MF, McQuillan JJ, Rosen GD, et al. Characterization of the promoter for vascular cell adhesion molecule-1 (VCAM-1). J Biol Chem 1991;267;16323-30.
- Whelan J. et al. An NF kappa B-like factor is essential but not sufficient for cytokine induction of endothelial leukocyte adhesion molecule 1 (ELAM-1) gene transcription. Nucleic Acids Res 1991;19:2645-2653. 3.
- Sharma HW, Higgins-Sochaski K, Perez JR, et al. A DNA motif present in alpha V integrin promoter exhibits dual binding preference to distinct transcription factors. Auticancer Res 1995;15:1857-1867. 33
 - Hansen SK, et al. A novel complex between the p65 subunit of NF-kappa B and c-Rel binds to a DNA element involved in the phorbol ester induction of the human urokinase gene. EMBO J 1992;11:205-213. 33,
- Sato H. Seiki M. Regulatory mechanism of 92 kDa type IV collagenase gene expression which is associated with invasiveness of tumor cells. Oncogene 1993;8:395-405. 34,
- Wang CY, Mayo MW, Korneluk R, et al. NF-kappaB antiapoptosis: induction of Grumont RJ. Rourke LJ, Gerondakis S. Rel-dependent induction of A1 transcription TRAFI and TRAF2 and c-1AP1 and c- 1AP2 to suppress caspase-8 activation. Science 1998;281:1680-1683. 35

36.

Jones PL, Ping D, Boss JM. Tumor necrosis factor alpha and interleukin-1 beta is required to protect B cells from antigen receptor ligation-induced apoptosis. Genes Dev 1999;13:400-411. 37.

regulate the murine manganese superoxide dismutase gene through a complex

- intronic enhancer involving C/EBP-beta and NF-kappaB. Mol Cell Biol 1997;17:
- Lee HH, Dadgostar H, Cheng Q, et al. NF-kappaB-mediated up-regulation of Bcl-x and Bfl-1/A1 is required for CD40 survival signaling in B lymphocytes. Proc Natl Acad Sci U S A 1999:96:9136-9141. 38
- Micheau O, Lens S, Gaide O. et al. NF-kappaB signals induce the expression of c-FLIP. Mol Cell Biol 2001;21:5299-5305. 39,
 - De Smaele E, et al. Induction of gadd45beta by NF-kappaB downregulates pro-apoptotic JNK signalling. Nature 2001;414:308-313. 40.
- Beg AA, Sha WC, Bronson RT, et al. Embryonic lethality and liver degeneration in mice lacking the RelA component of NF-kappa B. Nature 1995;376:167-170. ₩.
 - Li Q, Van Antwerp D, Mercurio F, et al. Severe liver degeneration in mice lacking the IkappaB kinase 2 gene. Science 1999;284:321-325. 42
- Tanaka M, et al. Embryonic lethality, liver degeneration, and impaired NF-kappa B activation in IKK-beta-deficient mice. Immunity 1999;10:421-429. 43.
- defects in proliferation, differentiation, germ-line CH transcription, and lg class Snapper CM, et al. B cells from p50/NF-kappa B knockout mice have selective switching. J Immunol 1996;156:183-191. 4.
 - Caamano JH, et al. Nuclear factor (NF)-kappa B2 (p100/p52) is required for normal splenic microarchitecture and B cell-nediated immune responses. J Exp Med 1998:187:185-196. 45.
- defects in humoral responses, germinal center reactions, and splenic microarchitecture. J Exp Med 1998;187:147-159. Franzoso G, et al. Mice deficient in nuclear factor (NF)-kappa B/p52 present with 46.
- Kontgen F. et al. Mice lacking the c-rel proto-oncogene exhibit defects in lymphocyte proliferation, humoral immunity, and interleukin-2 expression. Genes Dev 1995;9:1965-1977. 4.
 - lotsova V, et al. Osteopetrosis in mice lacking NF-kappaB1 and NF-kappaB2. Nat Med 1997;3:1285-1289. ₩.
 - Hennighausen L. Robinson GW. Signaling pathways in mammary gland development. Dev Cell 2001;1:467-475. 49.
- Hennighausen L, Robinson GW. Think globally, act locally: the making of a mouse mammary gland. Genes Dev 1998;12:449-455. 50.
 - Brantley DM, et al. Dynamic expression and activity of NF-kappaB during post-natal mammary gland morphogenesis. Mech Dev 2000:97:149-155. 3
 - Clarkson RW, Watson CJ. NF-kappaB and apoptosis in mammary epithelial cells. J Mammary Gland Biol Neoplasia 1999;4:165-175. 52.
- Clarkson RW, et al. NF-kappaB inhibits apoptosis in murine mammary epithelia. J Biol Chem 2000;275:12737-12742. 53
- paB/p50 in association with the growth and morphogenesis of normal and Brantley DM, et al. Nuclear factor-kappaB (NF-kappaB) regulates proliferation and Varela L.M., Stangle-Castor NC, Shoemaker SF, et al. TNFolpha induces NFkaptransformed rat mammary epithelial cells. J Cell Physiol 2001;188:120-131. 54. 55.
 - branching in mouse mammary epithelium. Mol Biol Cell 2001;12:1445-1455.
- Varela LM, Ip MM. Tumor necrosis factor-alpha: a multifunctional regulator of 56.
 - mammary gland development. Endocrinology 1996;137:4915-4924. Cao Y, et al. IKKalpha provides an essential link between RANK signaling and cyclin D1 expression during mammary gland development. Cell 2001;107:763-775. 57.

- Fata JE, et al. The oxteoclast differentiation factor oxteoprotegerin-ligand is essential for mammary gland development. Cell 2000;103:41-50. 38
 - Sicinski P. et al. Cyclin D1 provides a link between development and oncogenesis in the retina and breast. Cell 1995;82:621-630. 80
- Fantl V. Stamp G. Andrews A, et al. Mice lacking cyclin D1 are small and show defects in eye and mamnary gland development. Genes Dev 1995:9:2364-2372. 66
 - Duffy MJ, et al. Urokinase-plasminogen activator, a new and independent prognostic marker in breast cancer. Cancer Res 1990;50:6827-6829. 61:
- Nakshatri H. Bhat-Nakshatri P. Martin DA. et al. Constitutive activation of NF-kappaB during progression of breast cancer to hormone-independent growth. Mol Cell Biol 1997;17:3629-3639. 62.
- Newton TR, et al. Negative regulation of transactivation function but not DNA binding of NF-kappaB and AP-1 by IkappaBbeta1 in breast cancer cells. J Biol Chem 1999:274:18827-18835. 63
 - Sovak MA, et al. Aberrant nuclear factor-kappaB/Rel expression and the pathogenesis of breast cancer. J Clin Invest 1997;100:2952-2960. Ŧ.
- Cogswell PC, Guttridge DC, Funkhouser WK, et al. Selective activation of NF-kappaB subunits in human breast cancer; potential roles for NF-kappaB2/p52 and for Bel-3. Oncogene 2000;19:1123-1131. 65.
 - Dejardin E, et al. Highly-expressed p100/p52 (NFKB2) sequesters other NF-kappa B-related proteins in the cytoplasm of human breust cancer cells. Oncogene 1995;11:1835-1841. 99
- Stein B, et al. Cross-coupling of the NF-kappa B p65 and Fos/Jun transcription factors produces potentiated biological function. EMBO J 1993;12:3879-3891. 67.
- Platet N. Cunat S. Chalbos D. et al. Unliganded and liganded estrogen receptors protect against cancer invasion via different mechanisms. Mol Endocrinol 2000; . 98
- Bhat-Nakshatri P. Newton TR, Goulet R Jr, et al. NF-kappaB activation and interleukín 6 production in fibroblasts by estrogen receptor-negative breast cancer cell-derived interleukin Lalpha. Proc Natl Acad Sci U S A 1998;95:6971-6976. 66
- Bhat-Nakshatri P, Sweeney CJ, Nakshatri H. Identification of signal transduction pathways involved in constitutive NF-kappaB activation in breast cancer cells. Oncogene 2002;21:2066-2078. 3
- Biswas DK, Cruz AP, Gansberger E, et al. Epidermal growth factor-induced nuclear factor kappa B activation: a major pathway of cell-cycle progression in estrogen-receptor negative breast cancer cells. Proc Natl Acad Sci U S A 2000;97:8542-8547. Ξ:
- Biswas DK, et al. The nuclear factor kappa B (NF-kappa B): a potential therapeutic target for estrogen receptor negative breast cancers. Proc Natl Acad Sci U S A 2001;98:10386-10391. 72;
 - Nozaki S, Sledge GW Jr, Nakshatri H. Caucer cell-tlerived interleukin lalpha contributes to autocrine and paracrine induction of pro-metastatic genes in breast cancer. Biochem Biophys Res Commun 2000;275:60-62. 73,
- Lupu R, et al. The significance of heregulin in breast cancer tumor progression and drug resistance. Breast Cancer Res Treat 1996;38:57-66. 첫
- Slamon DJ, et al. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. Science 1987;235:177-182. 75.
- Bunyaratavej P, Hullinger TG, Somerman MJ. Bone morphogenetic proteins 76.

- secreted by breast cancer cells upregulate bone sialoprotein expression in preosteoblast cells. Exp Cell Res 2000;260:324-333.
- 77. Normanno N, Ciardiello F. EGF-related peptides in the pathophysiology of the mammary gland. J Mammary Gland Biol Neoplasia 1997;2:143-151.
- 78. Yee D, Rosen N, Favoni RE. et al. The insulin-like growth factors, their receptors, and their binding proteins in human breast cancer. Cancer Treat Res 1991;53:93-106.
- 79. Walker RA. The complexities of breast cancer desmoplasia. Breast Cancer Res 2001;3:143-145.
- 80. Dong G, Chen Z, Kato T, et al. The host environment promotes the constitutive activation of nuclear factor-kappaB and proinflammatory cytokine expression during metastatic tumor progression of murine squamous cell carcinoma. Cancer Res 1999;59:3495-3504.

,

- Romieu-Mourez R, et al. Roles of IKK kinases and protein kinase CK2 in activation of nuclear factor-kappaB in breast cancer. Cancer Res 2001;61:3810-3818.
- Patel NM, et al. Paclitaxel sensitivity of breast cancer cells with constitutively active NF-kappaB is enhanced by IkappaBalpha super-repressor and parthenolide. Oncogene 2000;19:4159-4169.
 - 83. Perou CM, et al. Molecular portraits of human breast tumours. Nature 2000;406: 747-752.
- 84. Perou CM, et al. Distinctive gene expression patterns in human mammary epithelial cells and breast cancers. Proc Natl Acad Sci U S A 1999;96:9212-9217.
- 85. Nozaki S, Sledge GW Jr, Nakshatri H. Repression of GADD153/CHOP by NF-kappaB: a possible cellular defense against endoplasmic reticulum stressinduced cell death. Oncogene 2001;20:2178-2185.
 - 86. Kim DW, et al. Activation of NF-kappaB/Rel occurs early during neoplastic transformation of mammary cells. Carcinogenesis 2000;21:871-879.
- 87. Jansen-Durr P. et al. Differential modulation of cyclin gene expression by MYC. Proc Natl Acad Sci U S A 1993;90:3685-3689.
- 88. Gillett C, et al. Amplification and overexpression of cyclin D1 in breast cancer detected by immunohistochemical staining. Cancer Res 1994;54:1812-1817.
- 89. Weinstat-Saslow D, et al. Overexpression of cyclin D mRNA distinguishes invasive and in situ breast carcinomas from non-malignant lesions. Nat Med 1995;1:1257-1260.
- Umekira Y, Ohi Y, Sagara Y, et al. Overexpression of cyclinD1 predicts for poor prognosis in estrogen receptor-negative breast cancer patients. Int J Cancer 2002;98:415-418.
- Yu Q, Geng Y, Sicinski P. Specific protection against breast cancers by cyclin D1 ablation. Nature 2001;411:1017-1021.
 - 92. Vousden KH, Lu X. Live or let die: the cell's response to p53. Nat Rev Cancer 2002;2:594-604.
- Ravi R, et al. p53-mediated repression of nuclear factor-kappaB RelA via the transcriptional integrator p500. Cancer Res 1998;58:4531-4536.
 - 94. Tergaonkar V, Pando M, Vafa O, et al. p53 stabilization is decreased upon NFkappaB activation: a role for NFkappaB in acquisition of resistance to chemotherapy. Cancer Cell 2002;1:493-503.

- Ryan KM, Ernst MK, Rice NR. et al. Role of NF-kappaB in p53-mediated. programmed cell death. Nature 2000;404:892-897.
 - Kaufman RJ. Stress signaling from the lumen of the endoplasmic reticulum: coordination of gene transcriptional and translational controls. Genes Dev 1999; 13:1211-1233.
- 97. Chilov D, et al. Genomic organization of human and mouse genes for vascular endothelial growth factor C, J Biol Chem 1997;272:25176-25183.
- 98. Dong G, et al. Molecular profiling of transformed and metastatic murine squamous carcinoma cells by differential display and cDNA microarray reveals altered expression of multiple genes related to growth, apoptosis, angiogenesis, and the NF-kappaB signal pathway. Cancer Res 2001:61:4797-4808.
- 99. Loukinova E, et al. Growth regulated oncogene-alpha expression by murine squamous cell carcinoma promotes tumor growth, metastasis, leukocyte infiltration and angiogenesis by a host CXC receptor-2 dependent mechanism. Oncogene 2000;19:3477-3486.
 - Grignani G, Maiolo A. Cytokines and hemostasis. Haematologica 2000;85:967-073
- 101. Chiarugi V, Ruggiero M, Magnelli L. Molecular polarity in endothelial cells and tumor-induced angiogenesis. Oncol Res 2000;12:1-4.
- 102. Manna SK, Zhang HJ. Yan T. et al. Overexpression of manganese superoxide dismutase suppresses tumor necrosis factor-induced apoptosis and activation of nuclear transcription factor-kappaB and activated protein-1. J Biol Chem 1998; 273:13245-13254.
- Delhalle S. Deregowski V, Benoit V, et al. NF-kappaB-dependent MnSOD expression protects adenocarcinoma cells from TNF-alpha-induced apoptosis. Oncogene 2002;21:3917-3924.
- 104. Toillon RA, et al. Normal breast epithelial cells induce apoptosis of breast cancer cells via Fas signaling. Exp Cell Res 2002;275:31-43.
 - Datta SR, Brunet A, Greenberg ME. Cellular survival: a play in three Akts. Genes Dev 1999;13:2905-2927.
- 106. Hurchinson J, Jin J. Cardiff R, et al. Activation of AKT (protein kinase B) in mammary epithelium provides a critical cell survival signal required for tumor progression. Mol Cell Biol 2002;21:2203-2212.
 - Meng F, Liu L, Chin PC, et al. AKT is a downstream target of NF-kappa B. J Biol Chem 2002;277:29674-29680.
 - 108. Bellacosa A, et al. Molecular alterations of the AKT2 oncogene in ovarian and breast carcinomas. Int J Cancer 1995;64:280-285.
- Nakatani K. et al. Up-regulation of Akt3 in estrogen receptor-deficient breast cancers and androgen-independent prostate cancer lines. J Biol Chem 1999;274: 21528-21532.
- 110. Vakkala M, et al. Inducible nitric oxide synthase expression, apoptosis, and angiogenesis in in situ and invasive breast carcinomas, Clin Cancer Res 2000:6: 2408-2416.
- Half E, et al. Cyclooxygenaso-2 expression in human breast cancers and adjacent ductal carcinoma in situ. Cancer Res 2002;62:1676-1681.
- Costa C, et al. Cyclo-oxygenase 2 expression is associated with angiogenesis and lymph node metastasis in human breast cancer. J Clin Pathol 2002;55:429-434.

- Ristimaki A, et al. Prognostic significance of clevated cyclooxygenase-2 expression in breast cancer. Cancer Res 2002;62:632-635. 133.
 - Hart IR, Saini A. Biology of tumour metastasis. Lancet 1992;339:1453-1457. 7,
- Edwards DR, Murphy G. Cancer: proteases: invasion and more. Nature 1998;394; 115.
- Kim J. Yu W, Kovalski K, et al. Requirement for specific proteases in cancer cell intravasation as revealed by a novel semiquantitative PCR-based assay. Cell 1998:94:353-362. 116,
- transformed and control fibroblasts extravasate equally well. Proc Natl Acad Sci U Koop S, et al. Independence of metastatic ability and extravasation: metastatic ras-S A 1996;93:11080-11084. 117.
- E. Reich, Ossowski L. Antibodies to plasminogen activator inhibit human tumor metastasis. Cell 1983;35:611-619. 18
- Sliva D, Rizzo MT. English D. Phosphatidylinositol 3-kinase and NF-kappaB regulate motility of invasive MDA-MB-231 human breast cancer cells by the secretion of urokinase-type plasminogen activator. J Biol Chem 2002:277:3150-119.
- Andela VB, Schwarz EM, Puzas JE, et al. Tumor metastasis and the reciprocal regulation of prometastatic and antimetastatic factors by nuclear factor kappaB. Cancer Res 2000;60:6557-6562. 120.
 - Martin TJ, Moseley JM. Mechanisms in the skeletal complications of breast cancer. Endocr Relat Cancer 2000;7:271-284. 2
- van Eys J. Nutrition and cancer: physiological interrelationships. Annu Rev Nutr 1985;5:435-461. 22.
 - Roodman GD. Biology of osteoclast activation in cancer. J Clin Oncol 2001;19: 3562-3571. 23.
- Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. Nat Rev Cancer 2002;2:584-593. 24.
- Mancino AT, Klimberg VS, Yamamoto M, et al. Breast cancer increases osteoclastogenesis by secreting M-CSF and upregulating RANKL in stromal cells. J Surg Res 2001;100:18-24. 25.
- Guise TA, et al. Evidence for a causal role of parathyroid hormone-related protein in the pathogenesis of human breast cancer-mediated osteolysis. J Clin Invest 1996:98:1544-1549. 26.
- Zimmers TA, et al. Induction of cachexia in mice by systemically administered myostatin. Science 2002;296:1486-1488. 127.
- Argiles JM, Lopez-Soriano FJ. The role of cytokines in cancer cachexia. Med Res Rev 1999;19:223-248. 28
- Tisdale MJ. Metabolic abnormalities in cachexía and anorexia. Nutrition 2000;16: 1013-1014. 129.
- Guttridge DC, Mayo MW, Madrid LV, et al. NF-kappaB-induced loss of MyoD messenger RNA: possible role in muscle decay and cachexia. Science 2000:289: 2363-2366. 130.
- Das KC, White CW. Activation of NF-kappaB by antineoplastic agents: role of protein kinase C. J Biol Chem 1997;272:14914-14920. 13
- Hwang S, Ding A. Activation of NF-kappa B in murine macrophages by taxol. Cancer Biochem Biophys 1995;14:265-272. 132.
- Curr Probl Cancer, September/October 2002

- Huang TT, et al. NF-kappaB activation by camptothecin: a linkage between nuclear DNA damage and cytoplasmic signaling events. J Biol Chem 2000:275:9501-9509. 333
 - Beg AA, Baltimore D. An essential role for NF-kappaB in preventing TNF-alphainduced cell death. Science 1996;274;782-784. ž
- Wang CY, Mayo MW, Baldwin AS Jr. TNF- and cancer therapy-induced apoptosis: potentiation by inhibition of NF-kappaB. Science 1996;274;784-787. 35.
- Van Antwerp DJ, Martin SJ. Kafri T. et al. Suppression of TNF-alpha-induced apoptosis by NF-kappaB. Science 1996;274:787-789. 36.
 - Green DR, Reed JC. Mitochondria and apoptosis. Science 1998:281:1309-1312. 337.
- Deveraux QL, Reed JC, IAP family proteins: suppressors of apoptosis. Genes Dev 1999;13:239-252.
 - Thomberry NA, Lazebnik Y. Caspases: enemies within, Science 1998;281:1312-39.
- Wang CY, Cusack JC Jr, Liu R, et al. Control of inducible chemoresistance: enhanced antí-tumor therapy through increused apoptosis by inhibition of NFkappaB. Nat Med 1999:5:412-417. 40.
- Hehner SP, et al. Sesquiterpene lactones specifically inhibit activation of NFkB by preventing the degradation of LkB- α and LkB- β . J Biol Chem 1998;273:1288-1297. 4
- Weldon CB, et al. NF-kappa B-mediated chemoresistance in breast cancer cells. Surgery 2001;130:143-150. <u>5</u>
 - Huang Y, Johnson KR, Norris J, et al. Nuclear factor-kappaB/IkappaB signaling pathway may contribute to the mediation of paclitaxel-induced apoptosis in solid tumor cells. Cancer Res 2000;60:4426-4432. 43.
- Auphan N, DiDonato JA. Rosette C, et al. Immunosuppression by glucocorticoids: inhibition of NF-kappa B activity through induction of I kappa B synthesis. Science 1995:270:286-290. 4.
- Scheinman RI. Cogswell PC, Lofquist AK, et al. Role of transcriptional activation of I kappa B alpha in mediation of immunosuppression by glucocorticoids. Science 145.
- Fukuda K, et al. Inhibition by berberine of cyclooxygenase-2 transcriptional activity in human colon cancer cells. J Ethnopharmacol 1999;66:227-233. 4
 - Habtemariam S. Cistifolin, an integrin-dependent cell adhesion blocker from the anti-rheumatic herbal drug, gravel root (rhizome of Eupatorium purpureum). Planta Med 1998;64:683-685. 47
 - Murphy JJ, Heptinstall S, Mitchell JR, Randomised double-blind placebo-controlled trial of feverfew in migraine prevention. Lancet 1988;2:189-192. φį
- Hehner SP, Hofmann TG, Droge W. et al. The antiinflammatory sesquiterpene lacrone parthenolide inhibits NF-kappa B by targeting the I kappa B kinase complex. J Immunol 1999;163;5617-5623. 49
 - Sobota R, Szwed M. Kasza A, et al. Parthenolide inhibits activation of signal transducers and activators of transcription (STATs) induced by cytokines of the IL-6 family. Biochem Biophys Res Commun 2000;267:329-333. 50
- proinflammatory cytokines by sesquiterpene lactones in macrophages correlates with the inhibition of MAP kinases. Biochem Biophys Res Commun 1996;226: Hwang D, et al. Inhibition of the expression of inducible cyclooxygenase and 151.
- Singh S, Aggarwal BB. Activation of transcription factor NF-kappa B is suppressed by curcumin (diferuloylmethane). J Biol Chem 1995;270:24995-25000. 152.

- Lin YL, Lin JK. (-)-Epigallocatechin-3-gallate blocks the induction of nitric oxide synthase by down-regulating lipopolysaccharide-induced activity of transcription factor nuclear factor-kappaB. Mol Pharmacol 1997;52:465-472.
 - 154. Pereira MA, et al. Effects of the phytochemicals, curcumin and quercetin, upon azoxymethane-induced colon cancer and 7,12-dimethylbenz anthracene-induced mammary cancer in rats. Carcinogenesis 1996;17;1305-1311 [a].
- 155. Rao CV, Rivenson A, Simi B, et al. Chemoprevention of colon carcinogenesis by dietary curcumin, a naturally occurring plant phenolic compound. Cancer Res 1995;57:549.266
- Kopp E, Ghosh S. Inhibition of NF-kappa B by sodium salicylate and aspirin. Science 1994;265:956-959.
 - Yin MJ, Yamamoto Y, Gaynor RB. The anti-inflammatory agents aspirin and salicylate inhibit the activity of I(kappa)B kinase-beta. Nature 1998;396:77-80.
 - Rossi A, et al. Anti-inflammatory cyclopentenone prostaglandins are direct inhibitors of IkappaB kinase. Nature 2000;403:103-108.
- 169. Chaturvedi MM, et al. Sanguinarine (pseudochelerythrine) is a potent inhibitor of NF-kappaB activation, IkappaBalpha phosphorylation, and degradation, J Biol Chem 1997;272:30129-30134.
 - Ji C, Kozak KR, Marnett LJ. IkappaB kinase, a molecular target for inhibition by 4-hydroxy-2-nonenal. J Biol Chem 2001;276:18223-18228.
- 161. May MJ, et al. Selective inhibition of NF-kappaB activation by a peptide that blocks the interaction of NEMO with the IkappaB kinase complex. Science 2000;289:1550-1554.
- Adams J. Development of the proteasome inhibitor PS-341. Oncologist 2002;7:9-16.
- 163. Shattuck-Brandt RL, Richmond A. Enhanced degradation of L-kappaB alpha contributes to endogenous activation of NF-kappaB in Hs294T melanoma cells. Cancer Res 1997;57:3032-3039.
- McKay LI, Cidlowski JA. Cross-talk between nuclear factor-kappa B and the steroid hormone receptors: mechanisms of mutual antagonism. Mol Endocrinol 1998;12:45-56.
- 165. Caldenhoven E, et al. Negative cross-talk between RelA and the glucocorticoid receptor: a possible mechanism for the antiinflammatory action of glucocorticoids. Mol Endocrinol 1995;9:401-412.
 - 166. Reichardt HM, et al. Repression of inflammatory responses in the absence of DNA binding by the glucocorticoid receptor. EMBO J 2001;20:7168-7173.
 - 167. Powles TJ, et al. Treatment of disseminated breast cancer with tamoxifen, aminoglutethimide, hydrocortisone, and danazol, used in combination or sequentially. Lancet 1984;1:1369-1373.
- 168. Cameron DA, et al. Primary systemic therapy for operable breast cancer: 10-year survival data after chemotherapy and hormone therapy. Br J Cancer 1997;76:1099-1105
- Wagstaff AJ, Goa KL. Rosiglitazone: a review of its use in the management of type 2 diabetes mellitus. Drugs 2002;62:1805-1837.
- 70. Mehta RG, Williamson E, Patel M, et al. A ligand of peroxisome proliferator-activated receptor gamma. retinoids, and prevention of preneoplastic mammary lesions. J Natl Cancer Inst 2000;92:418-423.
- 171. Badawi AF, Badr MZ. Chemoprevention of breast cancer by targeting cyclooxy-

308

- genase-2 and peroxisome proliferator-activated receptor-gamma. Int J Oncol 2002;20:1109-1122.
 - 172. Ricote M, Li AC, Willson TM, et al. The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. Nature 1998; 391-79,87
- 173. Jiang C, Ting AT, Seed B. PPAR-gumma agonísts inhibit production of monocyte inflammatory cytokines. Nature 1998;391:82-86.
 - 174. Patel L, et al. Tumor suppressor and anti-inflammatory actions of PPARyamma agonists are mediated via upregulation of PTEN. Curr Biol 2001;11:764-768.
- 175. Koul D. Yao Y, Abbruzzese JL, et al. Tumor suppressor MMAC/PTEN inhibits cytokine-induced NFkappaB activation without interfering with the IkappaB degradation pathway. J Biol Chem 2001;276:11402-11408.
- 176. Ozes ON, et al. NF-kappuB activation by tumour necrosis factor requires the AKT serine-threonine kinase. Nature 1999;401:82-85.
- 177. Clay CE, Atsumi GI, High KP, et al. Early de novo gene expression is required for 15-deoxy-Delta 12.14- prostaglandin J2-induced apoptosis in breast cancer cells. J Biol Chem 2001;276:47131-47135.
- 178. Higgins KA, et al. Antisense inhibition of the p65 subunit of NF-kappa B blocks unnorigenicity and causes tumor regression. Proc Natl Acad Sci U S A 1993;90:
- 179. Neurath MF, Pettersson S, Meyer zum Buschenfelde KH, et al. Local administration of antisense phosphorothioate oligonucleotides to the p65 subunit of NF-kappa B abrogates established experimental colitis in mice. Nat Med 1996;2:998-1004.
 - 180. Hannon GJ. RNA interference. Nature 2002;418:244-251.
- Lavon I, et al. High susceptibility to bacterial infection, but no liver dysfunction, in mice compromised for hepatocyte NF-kappaB activation. Nat Med 2000;6:573-577
- 182. Johnston SR, et al. Changes in estrogen receptor, progesterone receptor, and pS2 expression in tamoxifen-resistant human breast cancer. Cancer Res 1995;55:3331-3338.
 - Gu Z, et al. Association of interferon regulatory factor-1, nucleophosmin, nuclear factor-kappaB, and cyclic AMP response element binding with acquired resistance to Faslodex (ICT 182,780). Cancer Res 2002;62:3428-3437.